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(54) Title: NOVEL LIPASES AND USES THEREOF

(57) Abstract: The invention relates to a newly identified polynucleotide sequence comprising a gene that encodes a novel lipolytic enzyme from *Aspergillus niger*. The invention features the full length nucleotide sequence of the novel gene, the cDNA sequence comprising the full length coding sequence of the novel lipolytic enzyme as well as the amino acid sequence of the full-length functional protein and functional equivalents thereof. The invention also relates to methods of using these enzymes in industrial processes and methods of diagnosing fungal infections. Also included in the invention are cells transformed with a polynucleotide according to the invention and cells wherein a lipolytic enzyme according to the invention is genetically modified to enhance its activity and/or level of expression.

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NOVEL LIPASES AND USES THEREOF

Field of the invention

5 The invention relates to newly identified polynucleotide sequence comprising a gene that encodes a novel lipolytic enzyme from *Aspergillus niger*. The invention features the full length nucleotide sequence of the novel gene, the cDNA sequence comprising the full length coding sequence of the novel lipolytic enzyme as well as the amino acid sequence of the full-length lipolytic enzyme and functional equivalents thereof. The invention also relates to methods of using these enzymes in industrial processes and methods of diagnosing fungal infections. Also included in the invention are cells transformed with a polynucleotide according to the invention and cells wherein a lipolytic enzyme according to the invention is genetically modified to enhance its activity and/or level of expression.

15

Background of the invention

Baked products such as bread are prepared from a dough which is usually made from the basic ingredients (wheat) flour, water and optionally salt. Depending on the baked products, other ingredients added may be sugars, flavours etceteras. For leavened products, primarily baker's yeast is used next to chemical leavening systems such as a combination of an acid (generating compound) and bicarbonate.

In order to improve the handling properties of the dough and/or the final properties of the baked products there is a continuous effort to develop processing aids with improving properties. Processing aids are defined herein as compounds that improve the handling properties of the dough and/or the final properties of the baked products. Dough properties that may be improved comprise machineability, gas retaining capability, reduced stickness, elasticity, extensibility, moldability etcetera. Properties of the baked products that may be improved comprise loaf volume, crust crispiness, crumb texture and softness, flavour relative staleness and shelf life. These dough and/or baked product improving processing aids can be divided into two groups: chemical additives and enzymes (also referred to as baking enzymes).

Yeast, enzymes and chemical additives are generally added separately to the dough. Yeast may be added as a liquid suspension, in a compressed form or as

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active dry (ADY) or instant dry yeast (IDY). The difference between these yeast formulations is the water- and yeast dry matter content. Liquid yeast has a yeast dry matter content of less than 25% (w/v). Cream yeast is a particular form of liquid yeast and has a dry matter content between 17 and 23% (w/v). Compressed yeast has a yeast dry matter content between 25-35% (w/v) while the dry yeast formulations have a yeast dry matter content between 92-98% (w/v).

Enzymes may be added in a dry, e.g. granulated form or in liquid form. The chemical additives are in most cases added in powder form. Also, processing aid compositions which are tailored to specific baking applications, may be composed of a dedicated mixture of chemical additives and enzyme.

The preparation of a dough from the ingredients and processing aids described above is well known in the art and comprises mixing of said ingredients and processing aids and one or more moulding and fermentation steps.

The preparation of baked products from such doughs is also well known in the art and may comprise molding and shaping and further fermentation of the dough followed by baking at required temperatures and baking times.

Chemical additives with improving properties comprise oxidising agents such as ascorbic acid, bromate and azodicarbonate, reducing agents such as L-cysteine and glutathione, emulsifiers acting as dough conditioners such as diacetyl tartaric esters of mono/diglycerides (DATEM), sodium stearoyl lactylate (SSL) or calcium stearoyl lactylate (CSL), or acting as crumb softeners such as glycerol monostearate (GMS) etceteras, fatty materials such as triglycerides (fat) or lecithin and others.

As a result of a consumer-driven need to replace the chemical additives by more natural products, several baking enzymes have been developed with dough and/or baked product improving properties and which are used in all possible combinations depending on the specific baking application conditions. Suitable enzymes include starch degrading enzymes, arabinoxylan- and other hemicellulose degrading enzymes, cellulose degrading enzymes, oxidizing enzymes, fatty material splitting enzymes, protein degrading, modifying or crosslinking enzymes.

Starch degrading enzymes are for instance endo-acting enzymes such as alpha-amylase, maltogenic amylase, pullulanase or other debranching enzymes and exo-acting enzymes that cleave off glucose (amyloglucosidase), maltose (beta-amylase), maltotriose, maltotetraose and higher oligosaccharides.

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Arabinoxylan- and other hemicellulose degrading enzymes are for instance xylanases, pentosanases, hemicellulase, arabinofuranosidase, glucanase and others.

Cellulose degrading enzymes are for instance cellulase, cellobiohydrolase and beta-glucosidase.

5 Oxidizing enzymes are for instance glucose oxidase, hexose oxidase, pyranose oxidase, sulfhydryl oxidase, lipoxygenase, laccase, polyphenol oxidases and others.

Fatty material splitting enzymes are for instance lipolytic enzymes such as triacylglycerol lipases, phospholipases (such as A₁, A₂, B, C and D) and galactolipases.

10 Protein degrading, modifying or crosslinking enzymes are for instance endo-acting proteases (serine proteases, metalloproteases, aspartyl proteases, thiol proteases), exo-acting peptidases that cleave off one amino acid, or dipeptide, tripeptide etceteras from the N-terminal (aminopeptidases) or C-terminal (carboxypeptidases) ends of the polypeptide chain, asparagines or glutamine deamidating enzymes such as deamidase and peptidoglutaminase or crosslinking enzymes such as transglutaminase.

15 Baking enzymes may conveniently be produced in microorganisms. Microbial baking enzymes are available from a variety of sources; Bacillus spec. are a common source of bacterial enzymes, whereas fungal enzymes are commonly produced in Aspergillus spec.

20 Baking enzymes may be used in a manifold of baked goods. The term "baked goods" is herein defined as to comprise bread products such as tin bread, loaves of bread, French bread as well as rolls, cakes, pies, muffins, yeast raised and cake doughnuts and the like.

25 In the above processes, it is advantageous to use baking enzymes that are obtained by recombinant DNA techniques. Such recombinant enzymes have a number of advantages over their traditionally purified counterparts. Recombinant enzymes may be produced at a low cost price, high yield, free from contaminating agents like bacteria or viruses but also free from bacterial toxins or contaminating other enzyme activities.

Object of the invention

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It is an object of the invention to provide novel polynucleotides encoding novel lipolytic enzymes with improved properties. A further object is to provide naturally and recombinantly produced lipolytic enzymes as well as recombinant strains producing these. Also fusion polypeptides are part of the invention as well as methods of making

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and using the polynucleotides and polypeptides according to the invention.

It is also an object of the invention to provide novel lipolytic enzymes, which solve at least one of the above-mentioned problems or to provide novel lipolytic enzymes, which have one or more improved properties if used in dough and/or baked products, selected from the group of increased strength of the dough, increased elasticity of the dough, increased stability of the dough, reduced stickiness of the dough, improved extensibility of the dough, improved machineability of the dough, increased volume of the baked product, improved crumb structure of the baked product, improved softness of the baked product, improved flavour of the baked product, improved anti-staling of the baked product, improved colour of the baked product, improved crust of the baked product or which have a broad substrate specificity.

Summary of the invention

15 The invention provides for novel polynucleotides encoding novel lipolytic enzymes. More in particular, the invention provides for polynucleotides having a nucleotide sequence that hybridises preferably under highly stringent conditions to a sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38. Consequently, the
20 invention provides nucleic acids that are more than 40% such as about 60%, preferably 65%, more preferably 70%, even more preferably 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% homologous to the sequences selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38.

25 In a more preferred embodiment the invention provides for such an isolated polynucleotide obtainable from a filamentous fungus, in particular *Aspergillus niger* is preferred.

In one embodiment, the invention provides for an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide with an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24,
30 27, 30, 33, 36 and 39 or functional equivalents thereof.

In a further preferred embodiment, the invention provides an isolated polynucleotide encoding at least one functional domain of a polypeptide selected from

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the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.

In a preferred embodiment the invention provides a lipolytic enzyme gene selected from the group consisting of SEQ ID NO: 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 5 34 and 37. In another aspect the invention provides a polynucleotide, preferably a cDNA encoding an *Aspergillus niger* lipolytic enzyme whose amino acid sequence is selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or variants or fragments of that polypeptide. In a preferred embodiment the cDNA has a sequence selected from the group consisting of SEQ ID NO: 2, 5, 8, 11, 14, 17, 20, 23, 10 26, 29, 32, 35 and 38 or functional equivalents thereof.

In an even further preferred embodiment, the invention provides for a polynucleotide comprising the coding sequence of the polynucleotides according to the invention, preferred is the polynucleotide sequence selected from the group consisting of SEQ ID NO: 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35 and 38.

15 The invention also relates to vectors comprising a polynucleotide sequence according to the invention and primers, probes and fragments that may be used to amplify or detect the DNA according to the invention.

In a further preferred embodiment, a vector is provided wherein the polynucleotide sequence according to the invention is functionally linked with regulatory 20 sequences suitable for expression of the encoded amino acid sequence in a suitable host cell, such as *Aspergillus niger* or *Aspergillus oryzae*. The invention also provides methods for preparing polynucleotides and vectors according to the invention.

The invention also relates to recombinantly produced host cells that contain heterologous or homologous polynucleotides according to the invention.

25 In another embodiment, the invention provides recombinant host cells wherein the expression of a lipolytic enzyme according to the invention is significantly increased or wherein the activity of the lipolytic enzyme is increased.

In another embodiment the invention provides for a recombinantly produced host cell that contains heterologous or homologous polynucleotide according to the 30 invention and wherein the cell is capable of producing a functional lipolytic enzyme according to the invention, preferably a cell capable of over-expressing the lipolytic enzyme according to the invention, for example an *Aspergillus* strain comprising an increased copy number of a gene or cDNA according to the invention.

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In yet another aspect of the invention, a purified polypeptide is provided. The polypeptides according to the invention include the polypeptides encoded by the polynucleotides according to the invention. Especially preferred is a polypeptide selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.

Accordingly, in one aspect the present invention provides a lipolytic enzyme composition containing as an active ingredient an enzyme selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.

In another aspect, the invention provides a method of making baked goods wherein there is incorporated into the dough used for making the baked goods one or more enzymes selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.

Fusion proteins comprising a polypeptide according to the invention are also within the scope of the invention. The invention also provides methods of making the polypeptides according to the invention.

The invention also relates to the use of the lipolytic enzyme according to the invention in any industrial process as described herein.

Detailed description of the invention

A lipolytic enzyme is defined herein as an enzyme exhibiting at least one and preferably two or three or four or more of the following lipolytic activities: triacylglycerol lipase, phospholipase A₁, phospholipase A₂, phospholipase B, phospholipase C, phospholipase D, lysophospholipase and galactolipase.

Polynucleotides

The present invention provides polynucleotides encoding lipolytic enzymes having an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof. The sequences of the seven genes encoding the lipolytic enzymes NBE028, NBE029, NBE030, NBE031, NBE032, NBE033, NBE034, NBE036, NBE038, NBE039, NBE043, NBE045 and NBE042 respectively were determined by sequencing genomic clones

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obtained from *Aspergillus niger*. The invention provides polynucleotide sequences comprising the genes encoding the lipolytic enzymes NBE028, NBE029, NBE030, NBE031, NBE032, NBE033, NBE034, NBE036, NBE038, NBE039, NBE043, NBE045 and NBE042 as well as their complete cDNA sequences and their coding sequences

5 (Table 1). Accordingly, the invention relates to isolated polynucleotides comprising the nucleotide sequences selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 or functional equivalents thereof.

10 Table 1.

lipolytic enzyme	Sequence (SEQ ID NO)		
	genomic	cDNA	amino acid
NBE028	1	2	3
NBE029	4	5	6
NBE030	7	8	9
NBE031	10	11	12
NBE032	13	14	15
NBE033	16	17	18
NBE034	19	20	21
NBE036	22	23	24
NBE038	25	26	27
NBE039	28	29	30
NBE043	31	32	33
NBE045	34	35	36
NBE042	37	38	39

More in particular, the invention relates to an isolated polynucleotide hybridisable under stringent conditions, preferably under highly stringent conditions, to a polynucleotide selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11,

15 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38. Advantageously, such polynucleotides may be obtained from filamentous fungi, in particular from *Aspergillus niger*. More specifically, the invention relates to an isolated polynucleotide having a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4,



5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38.

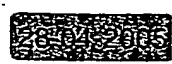
The invention also relates to an isolated polynucleotide encoding at least one functional domain of a polypeptide having an amino acid sequences selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.

As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which may be isolated from chromosomal DNA, which include an open reading frame encoding a protein, e.g. an *Aspergillus niger* lipolytic enzyme. A gene may include coding sequences, non-coding sequences, introns and regulatory sequences. Moreover, a gene refers to an isolated nucleic acid molecule as defined herein.

A nucleic acid molecule of the present invention, such as a nucleic acid molecule having the nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 or a functional equivalent thereof, can be isolated using standard molecular biology techniques and the sequence information provided herein. For example, using all or portion of the nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 as a hybridization probe, nucleic acid molecules according to the invention can be isolated using standard hybridization and cloning techniques (e. g., as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning: A Laboratory Manual.2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

Moreover, a nucleic acid molecule encompassing all or a portion of the nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence information contained in the nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis.



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Furthermore, oligonucleotides corresponding to or hybridisable to nucleotide sequences according to the invention can be prepared by standard synthetic techniques, e. g., using an automated DNA synthesizer.

In one preferred embodiment, an isolated nucleic acid molecule of the invention
5 comprises the nucleotide sequence shown in SEQ ID NO: 2. The sequence of SEQ ID NO: 2 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 1. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE028 polypeptide as shown in SEQ ID NO: 3.

In a second preferred embodiment, an isolated nucleic acid molecule of the
10 invention comprises the nucleotide sequence shown in SEQ ID NO: 5. The sequence of SEQ ID NO: 5 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 4. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE029 polypeptide as shown in SEQ ID NO: 6.

In a third preferred embodiment, an isolated nucleic acid molecule of the
15 invention comprises the nucleotide sequence shown in SEQ ID NO: 8. The sequence of SEQ ID NO: 8 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 7. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE030 polypeptide as shown in SEQ ID NO: 9.

In a fourth preferred embodiment, an isolated nucleic acid molecule of the
20 invention comprises the nucleotide sequence shown in SEQ ID NO: 11. The sequence of SEQ ID NO: 11 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 10. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE031 polypeptide as shown in SEQ ID NO: 12.

In a fifth preferred embodiment, an isolated nucleic acid molecule of the
25 invention comprises the nucleotide sequence shown in SEQ ID NO: 14. The sequence of SEQ ID NO: 14 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 13. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE032 polypeptide as shown in SEQ ID NO: 15.

In a sixth preferred embodiment, an isolated nucleic acid molecule of the
30 invention comprises the nucleotide sequence shown in SEQ ID NO: 17. The sequence of SEQ ID NO: 17 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 16. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE033 polypeptide as shown in SEQ ID NO: 18.

In a seventh preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 20. The sequence of SEQ ID NO: 20 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 19. This cDNA comprises the sequence encoding the

5 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 21.

In a eight preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 23. The sequence of SEQ ID NO: 23 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 22. This cDNA comprises the sequence encoding the

10 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 24.

In a ninth preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 26. The sequence of SEQ ID NO: 26 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 25. This cDNA comprises the sequence encoding the

15 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 27.

In a tenth preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 29. The sequence of SEQ ID NO: 29 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 28. This cDNA comprises the sequence encoding the

20 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 30.

In a eleventh preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 32. The sequence of SEQ ID NO: 32 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 31. This cDNA comprises the sequence encoding the

25 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 33.

In a twelfth preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 35. The sequence of SEQ ID NO: 35 corresponds to the coding region of the *Aspergillus niger* gene provided in SEQ ID NO: 34. This cDNA comprises the sequence encoding the

30 *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 36.

In a thirteenth preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO: 38. The sequence of SEQ ID NO: 38 corresponds to the coding region of the *Aspergillus niger* gene

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provided in SEQ ID NO: 37. This cDNA comprises the sequence encoding the *Aspergillus niger* NBE034 polypeptide as shown in SEQ ID NO: 39.

In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 or a functional equivalent of these nucleotide sequences. A nucleic acid molecule, which is complementary to another nucleotide sequence, is one that is sufficiently complementary to the other nucleotide sequence such that it can hybridize to the other nucleotide sequence thereby forming a stable duplex.

One aspect of the invention pertains to isolated nucleic acid molecules that encode a polypeptide of the invention or a functional equivalent thereof such as a biologically active fragment or domain, as well as nucleic acid molecules sufficient for use as hybridisation probes to identify nucleic acid molecules encoding a polypeptide of the invention and fragments of such nucleic acid molecules suitable for use as PCR primers for the amplification or mutation of nucleic acid molecules. An "isolated polynucleotide" or "isolated nucleic acid" is a DNA or RNA that is not immediately contiguous with both of the coding sequences with which it is immediately contiguous (one on the 5' end and one on the 3' end) in the naturally occurring genome of the organism from which it is derived. Thus, in one embodiment, an isolated nucleic acid includes some or all of the 5' non-coding (e.g., promoter) sequences that are immediately contiguous to the coding sequence. The term therefore includes, for example, a recombinant DNA that is incorporated into a vector, into an autonomously replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (e.g., a cDNA or a genomic DNA fragment produced by PCR or restriction endonuclease treatment) independent of other sequences. It also includes a recombinant DNA that is part of a hybrid gene encoding an additional polypeptide that is substantially free of cellular material, viral material, or culture medium (when produced by recombinant DNA techniques), or chemical precursors or other chemicals (when chemically synthesized). Moreover, an "isolated nucleic acid fragment" is a nucleic acid fragment that is not naturally occurring as a fragment and would not be found in the natural state.

As used herein, the terms "polynucleotide" or "nucleic acid molecule" are

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intended to include DNA molecules (e.g., cDNA or genomic DNA) and RNA molecules (e.g., mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA. The nucleic acid may be synthesized using oligonucleotide
5 analogs or derivatives (e.g., inosine or phosphorothioate nucleotides). Such oligonucleotides can be used, for example, to prepare nucleic acids that have altered base-pairing abilities or increased resistance to nucleases.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to a nucleic acid molecule according to the invention. Also included
10 within the scope of the invention are the complement strands of the nucleic acid molecules described herein.

Sequencing errors

15 The sequence information as provided herein should not be so narrowly construed as to require inclusion of erroneously identified bases. The specific sequences disclosed herein can be readily used to isolate the complete gene from filamentous fungi, in particular *Aspergillus niger* which in turn can easily be subjected to further sequence analyses thereby identifying sequencing errors.

20 Unless otherwise indicated, all nucleotide sequences determined by sequencing a DNA molecule herein were determined using an automated DNA sequencer and all amino acid sequences of polypeptides encoded by DNA molecules determined herein were predicted by translation of a DNA sequence determined as above. Therefore, as is
known in the art for any DNA sequence determined by this automated approach, any
25 nucleotide sequence determined herein may contain some errors. Nucleotide sequences determined by automation are typically at least about 90% identical, more typically at least about 95% to at least about 99.9% identical to the actual nucleotide sequence of the sequenced DNA molecule. The actual sequence can be more precisely determined by other approaches including manual DNA sequencing methods well known in the art.
30 As is also known in the art, a single insertion or deletion in a determined nucleotide sequence compared to the actual sequence will cause a frame shift in translation of the nucleotide sequence such that the predicted amino acid sequence encoded by a determined nucleotide sequence will be completely different from the amino acid

sequence actually encoded by the sequenced DNA molecule, beginning at the point of such an insertion or deletion.

The person skilled in the art is capable of identifying such erroneously identified bases and knows how to correct for such errors.

5

Nucleic acid fragments, probes and primers

A nucleic acid molecule according to the invention may comprise only a portion or a fragment of the nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38, for example a fragment which can be used as a probe or primer or a fragment encoding a portion of protein according to the invention. The nucleotide sequence determined from the cloning of the lipolytic enzyme gene and cDNA allows for the generation of probes and primers designed for use in identifying and/or cloning other lipolytic enzyme family members, as well as lipolytic enzyme homologues from other species. The probe/primer typically comprises substantially purified oligonucleotide which typically comprises a region of nucleotide sequence that hybridizes preferably under highly stringent conditions to at least about 12 or 15, preferably about 18 or 20, preferably about 22 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, or 75 or more consecutive nucleotides of a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 or of a functional equivalent thereof.

Probes based on the nucleotide sequences provided herein can be used to detect transcripts or genomic sequences encoding the same or homologous proteins for instance in other organisms. In preferred embodiments, the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme cofactor. Such probes can also be used as part of a diagnostic test kit for identifying cells that express a lipolytic enzyme protein.

30

Identity & homology

The terms "homology" or "percent identity" are used interchangeably herein. For the purpose of this invention, it is defined here that in order to determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the

sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the sequence of a first amino acid or nucleic acid sequence for optimal alignment with a second amino or nucleic acid sequence). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences (i.e., % identity = number of identical positions/total number of positions (i.e. overlapping positions) x 100). Preferably, the two sequences are the same length.

The skilled person will be aware of the fact that several different computer programmes are available to determine the homology between two sequences. For instance, a comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a preferred embodiment, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch (J. Mol. Biol. (48): 444-453 (1970)) algorithm which has been incorporated into the GAP program in the GCG software package (available at <http://www.gcg.com>), using either a Blossom 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. The skilled person will appreciate that all these different parameters will yield slightly different results but that the overall percentage identity of two sequences is not significantly altered when using different algorithms.

In yet another embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at <http://www.gcg.com>), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. In another embodiment, the percent identity two amino acid or nucleotide sequence is determined using the algorithm of E. Meyers and W. Miller (CABIOS, 4:11-17 (1989) which has been incorporated into the ALIGN program (version 2.0) (available at <http://vega.lgh.cnrs.fr/bin/align-guess.cgi>) using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The nucleic acid and protein sequences of the present invention can further be used as a "query sequence" to perform a search against public databases to, for

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example, identify other family members or related sequences. Such searches can be performed using the BLASTN and BLASTX programs (version 2.0) of Altschul, et al. (1990) J. Mol. Biol. 215:403—10. BLAST nucleotide searches can be performed with the BLASTN program, score = 100, word length = 12 to obtain nucleotide sequences
5 homologous to PLP03 nucleic acid molecules of the invention. BLAST protein searches can be performed with the BLASTX program, score = 50, word length = 3 to obtain amino acid sequences homologous to PLP03 protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., (1997) Nucleic Acids Res. 25(17):3389-3402. When utilizing
10 BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., BLASTX and BLASTN) can be used. See <http://www.ncbi.nlm.nih.gov>.

Hybridisation

15

As used herein, the term "hybridizing" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least about 50%, at least about 60%, at least about 70%, more preferably at least about 80%, even more preferably at least about 85% to 90%, more preferably at least 95% homologous to each
20 other typically remain hybridized to each other.

20

A preferred, non-limiting example of such hybridization conditions are hybridization in 6x sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 1x SSC, 0.1 % SDS at 50°C, preferably at 55°C, preferably at 60°C and even more preferably at 65°C.

25

Highly stringent conditions include, for example, hybridizing at 68°C in 5x SSC/5x Denhardt's solution / 1.0% SDS and washing in 0.2x SSC/0.1% SDS at room temperature. Alternatively, washing may be performed at 42°C.

The skilled artisan will know which conditions to apply for stringent and highly stringent hybridisation conditions. Additional guidance regarding such conditions is readily available in the art, for example, in Sambrook et al., 1989, Molecular Cloning, A
30. Laboratory Manual, Cold Spring Harbor Press, N.Y.; and Ausubel et al. (eds.), 1995, Current Protocols in Molecular Biology, (John Wiley & Sons, N.Y.).

Of course, a polynucleotide which hybridizes only to a poly A sequence (such

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as the 3' terminal poly(A) tract of mRNAs), or to a complementary stretch of T (or U) residues, would not be included in a polynucleotide of the invention used to specifically hybridize to a portion of a nucleic acid of the invention, since such a polynucleotide would hybridize to any nucleic acid molecule containing a poly (A) stretch or the complement thereof (e.g., practically any double-stranded cDNA clone).

Obtaining full length DNA from other organisms

In a typical approach, cDNA libraries constructed from other organisms, e.g. filamentous fungi, in particular from the species *Aspergillus* can be screened.

For example, *Aspergillus* strains can be screened for homologous polynucleotides by Northern blot analysis. Upon detection of transcripts homologous to polynucleotides according to the invention, cDNA libraries can be constructed from RNA isolated from the appropriate strain, utilizing standard techniques well known to those of skill in the art. Alternatively, a total genomic DNA library can be screened using a probe hybridisable to a polynucleotide according to the invention.

Homologous gene sequences can be isolated, for example, by performing PCR using two degenerate oligonucleotide primer pools designed on the basis of nucleotide sequences as taught herein.

The template for the reaction can be cDNA obtained by reverse transcription of mRNA prepared from strains known or suspected to express a polynucleotide according to the invention. The PCR product can be subcloned and sequenced to ensure that the amplified sequences represent the sequences of a new PLP03 nucleic acid sequence, or a functional equivalent thereof.

The PCR fragment can then be used to isolate a full length cDNA clone by a variety of known methods. For example, the amplified fragment can be labeled and used to screen a bacteriophage or cosmid cDNA library. Alternatively, the labeled fragment can be used to screen a genomic library.

PCR technology can also be used to isolate full-length cDNA sequences from other organisms. For example, RNA can be isolated, following standard procedures, from an appropriate cellular or tissue source. A reverse transcription reaction can be performed on the RNA using an oligonucleotide primer specific for the most 5' end of the amplified fragment for the priming of first strand synthesis.

The resulting RNA/DNA hybrid can then be "tailed" (e.g., with guanines) using a

standard terminal transferase reaction, the hybrid can be digested with RNase H, and second strand synthesis can then be primed (e.g., with a poly-C primer). Thus, cDNA sequences upstream of the amplified fragment can easily be isolated. For a review of useful cloning strategies, see e.g., Sambrook et al., supra; and Ausubel et al., supra.

5

Vectors

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding a protein according to the invention or a functional equivalent thereof. As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. The terms "plasmid" and "vector" can be used interchangeably herein as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vector includes one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operatively linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for

expression of the nucleotide sequence (e.g., in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signal). Such regulatory sequences are described, for example, in Goeddel; *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive or inducible expression of a nucleotide sequence in many types of host cells and those which direct expression of the nucleotide sequence only in a certain host cell (e.g. tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, encoded by nucleic acids as described herein (e.g. lipolytic enzymes, mutant lipolytic enzymes, fragments thereof, variants or functional equivalents thereof, fusion proteins, etc.).

The recombinant expression vectors of the invention can be designed for expression of lipolytic enzymes in prokaryotic or eukaryotic cells. For example, a protein according to the invention can be expressed in bacterial cells such as *E. coli* and *Bacillus species*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression vectors useful in the present invention include chromosomal-, episomal- and virus-derived vectors e.g., vectors derived from bacterial plasmids, bacteriophage, yeast episome, yeast chromosomal elements, viruses such as baculoviruses, papova viruses, vaccinia viruses, adenoviruses, fowl pox viruses, pseudorabies viruses and retroviruses, and vectors derived from combinations thereof, such as those derived from plasmid and bacteriophage genetic elements, such as cosmids and phagemids.

The DNA insert should be operatively linked to an appropriate promoter, such as the phage lambda PL promoter, the *E. coli* lac, trp and tac promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name a few. The skilled person

will know other suitable promoters. In a specific embodiment, promoters are preferred that are capable of directing a high expression level of lipolytic enzymes in filamentous fungi. Such promoters are known in the art. The expression constructs may contain sites for transcription initiation, termination, and, in the transcribed region, a ribosome binding site for translation. The coding portion of the mature transcripts expressed by the constructs will include a translation initiating AUG at the beginning and a termination codon appropriately positioned at the end of the polypeptide to be translated.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, transduction, infection, lipofection, cationic lipidmediated transfection or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (*Molecular Cloning: A Laboratory Manual, 2nd ed.* Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), Davis et al., *Basic Methods in Molecular Biology* (1986) and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methatrexate. A nucleic acid encoding a selectable marker is preferably introduced into a host cell on the same vector as that encoding a protein according to the invention or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g. cells that have incorporated the selectable marker gene will survive, while the other cells die).

Expression of proteins in prokaryotes is often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, e.g. to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to

increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety after purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

As indicated, the expression vectors will preferably contain selectable markers. Such markers include dihydrofolate reductase or neomycin resistance for eukaryotic cell culture and tetracycline or ampicillin resistance for culturing in *E. coli* and other bacteria. Representative examples of appropriate host include bacterial cells, such as *E. coli*, *Streptomyces* and *Salmonella typhimurium*; fungal cells, such as yeast; insect cells such as *Drosophila* S2 and *Spodoptera* Sf9; animal cells such as CHO, COS and Bowes melanoma; and plant cells. Appropriate culture mediums and conditions for the above-described host cells are known in the art.

Among vectors preferred for use in bacteria are pQE70, pQE80 and pQE-9, available from Qiagen; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16A, pNH18A, pNH46A, available from Stratagene; and ptc99a, pKK223-3, pKK233-3, pDR540, pRIT5 available from Pharmacia. Among preferred eukaryotic vectors are PWLNEO, pSV2CAT, pOG44, pZT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. Other suitable vectors will be readily apparent to the skilled artisan.

Known bacterial promoters for use in the present invention include *E. coli* lacI and lacZ promoters, the T3 and T7 promoters, the gpt promoter, the lambda PR, PL promoters and the trp promoter, the HSV thymidine kinase promoter, the early and late SV40 promoters, the promoters of retroviral LTRs, such as those of the Rous sarcoma virus ("RSV"), and metallothionein promoters, such as the mouse metallothionein-I promoter.

Inserting an enhancer sequence into the vector may increase transcription of the DNA encoding the polypeptides of the present invention by higher eukaryotes. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp that act to increase transcriptional activity of a promoter in a given host cell-type. Examples of enhancers include the SV40 enhancer, which is located on the late side of the replication origin at bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma

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enhancer on the late side of the replication origin, and adenovirus enhancers.

For secretion of the translated protein into the lumen of the endoplasmic reticulum, into the periplasmic space or into the extracellular environment, appropriate secretion signal may be incorporated into the expressed polypeptide. The signals may
5 be endogenous to the polypeptide or they may be heterologous signals.

The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals but also additional heterologous functional regions. Thus, for instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and
10 persistence in the host cell, during purification or during subsequent handling and storage. Also, peptide moieties may be added to the polypeptide to facilitate purification.

Polypeptides according to the invention

15 The invention provides an isolated polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39, an amino acid sequence obtainable by expressing the polynucleotide selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17,
20 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38 in an appropriate host. Also, a peptide or polypeptide comprising a functional equivalent of the above polypeptides is comprised within the present invention. The above polypeptides are collectively comprised in the term "polypeptides according to the invention"

The terms "peptide" and "oligopeptide" are considered synonymous (as is commonly recognized) and each term can be used interchangeably as the context
25 requires to indicate a chain of at least two amino acids coupled by peptidyl linkages. The word "polypeptide" is used herein for chains containing more than seven amino acid residues. All oligopeptide and polypeptide formulas or sequences herein are written from left to right and in the direction from amino terminus to carboxy terminus. The one-letter code of amino acids used herein is commonly known in the art and can be found in
30 Sambrook, et al. (*Molecular Cloning: A Laboratory Manual*, 2nd, ed. Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989)

By "isolated" polypeptide or protein is intended a polypeptide or protein removed from its native environment. For example, recombinantly produced

polypeptides and proteins expressed in host cells are considered isolated for the purpose of the invention as are native or recombinant polypeptides which have been substantially purified by any suitable technique such as, for example, the single-step purification method disclosed in Smith and Johnson, Gene 67:31-40 (1988).

5 The lipolytic enzyme according to the invention can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Most
10 preferably, high performance liquid chromatography ("HPLC") is employed for purification.

Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast,
15 higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes.

20 A lipolytic enzyme according to the invention may be advantageously used in baking processes. The amount of enzyme to be added to the dough is determined empirically. It may depend on the quality of the flour used, the degree of improvement which is required, the kind of bread or baked goods, the method of preparing the dough, the proportion of other ingredients etcetera.

25

Protein fragments

The invention also features biologically active fragments of the polypeptides according to the invention.

30 Biologically active fragments of a polypeptide of the invention include polypeptides comprising amino acid sequences sufficiently identical to or derived from the amino acid sequence of the lipolytic enzyme (e.g., the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and

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39), which include fewer amino acids than the full length protein, and exhibit at least one biological activity of the corresponding full-length protein. Typically, biologically active fragments comprise a domain or motif with at least one activity of the corresponding full length protein.

5 A biologically active fragment of a protein of the invention can be a polypeptide which is, for example, 10, 25, 50, 100 or more amino acids in length. Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the biological activities of the native form of a polypeptide of the invention.

10 The invention also features nucleic acid fragments which encode the above biologically active fragments of the lipolytic enzyme protein.

Fusion proteins

15 The proteins of the present invention or functional equivalents thereof, e.g., biologically active portions thereof, can be operatively linked to a non-lipolytic enzyme polypeptide (e.g., heterologous amino acid sequences) to form fusion proteins. As used herein, a lipolytic enzyme "chimeric protein" or "fusion protein" comprises a lipolytic enzyme polypeptide operatively linked to a non-lipolytic enzyme polypeptide. A "lipolytic enzyme polypeptide" refers to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 20 39, whereas a "non-lipolytic enzyme polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the lipolytic enzyme, e.g., a protein which is different from the lipolytic enzyme and 25 which is derived from the same or a different organism. Within a lipolytic enzyme fusion protein the lipolytic enzyme polypeptide can correspond to all or a portion of a lipolytic enzyme protein. In a preferred embodiment, a lipolytic enzyme fusion protein comprises at least one biologically active fragment of a lipolytic enzyme protein. In another preferred embodiment, a lipolytic enzyme fusion protein comprises at least two 30 biologically active portions of a lipolytic enzyme protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the lipolytic enzyme polypeptide and the non-lipolytic enzyme polypeptide are fused in-frame to each other. The non-lipolytic enzyme polypeptide can be fused to the N-terminus or C-terminus of the lipolytic enzyme

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polypeptide.

For example, in one embodiment, the fusion protein is a GST-lipolytic enzyme fusion protein in which the lipolytic enzyme sequence is fused to the C-terminus of the GST sequence. Such fusion proteins can facilitate the purification of recombinant lipolytic enzyme(s). In another embodiment, the fusion protein is a lipolytic enzyme protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian and Yeast host cells), expression and/or secretion of lipolytic enzyme can be increased through use of a heterologous signal sequence.

In another example, the gp67 secretory sequence of the baculovirus envelope protein can be used as a heterologous signal sequence (*Current Protocols in Molecular Biology*, Ausubel et al., eds., John Wiley & Sons, 1992). Other examples of eukaryotic heterologous signal sequences include the secretory sequences of melittin and human placental alkaline phosphatase (Stratagene; La Jolla, California). In yet another example, useful prokaryotic heterologous signal sequences include the phoA secretory signal (Sambrook et al., *supra*) and the protein A secretory signal (Pharmacia Biotech; Piscataway, New Jersey).

A signal sequence can be used to facilitate secretion and isolation of a protein or polypeptide of the invention. Signal sequences are typically characterized by a core of hydrophobic amino acids which are generally cleaved from the mature protein during secretion in one or more cleavage events. Such signal peptides contain processing sites that allow cleavage of the signal sequence from the mature proteins as they pass through the secretory pathway. The signal sequence directs secretion of the protein, such as from a eukaryotic host into which the expression vector is transformed, and the signal sequence is subsequently or concurrently cleaved. The protein can then be readily purified from the extracellular medium by art recognized methods. Alternatively, the signal sequence can be linked to the protein of interest using a sequence which facilitates purification, such as with a GST domain. Thus, for instance, the sequence encoding the polypeptide may be fused to a marker sequence, such as a sequence encoding a peptide, which facilitates purification of the fused polypeptide. In certain preferred embodiments of this aspect of the invention, the marker sequence is a hexahistidine peptide, such as the tag provided in a pQE vector (Qiagen, Inc.), among others, many of which are commercially available. As described in *Gentz et al, Proc. Natl. Acad. Sci. USA 86:821-824 (1989)*, for instance, hexahistidine provides for convenient

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purification of the fusion protein. The HA tag is another peptide useful for purification which corresponds to an epitope derived of influenza hemagglutinin protein, which has been described by Wilson *et al.*, *Cell* 37:767 (1984), for instance.

Preferably, a chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel *et al.* John Wiley & Sons: 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (e.g. a GST polypeptide). A lipolytic enzyme-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the lipolytic enzyme protein.

20

Functional equivalents

The terms "functional equivalents" and "functional variants" are used interchangeably herein. Functional equivalents of lipolytic enzyme encoding DNA are isolated DNA fragments that encode a polypeptide that exhibits a particular function of the *Aspergillus niger* lipolytic enzyme as defined herein. A functional equivalent of a lipolytic enzyme polypeptide according to the invention is a polypeptide that exhibits at least one function of an *Aspergillus niger* lipolytic enzyme as defined herein. Functional equivalents therefore also encompass biologically active fragments.

Functional protein or polypeptide equivalents may contain only conservative substitutions of one or more amino acids in the amino acid sequences selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or substitutions, insertions or deletions of non-essential amino acids. Accordingly, a non-

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essential amino acid is a residue that can be altered in the amino acid sequences selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 without substantially altering the biological function. For example, amino acid residues that are conserved among the lipolytic enzyme proteins of the present invention, are predicted to be particularly unamenable to alteration. Furthermore, amino acids conserved among the lipolytic enzyme proteins according to the present invention and other lipolytic enzymes are not likely to be amenable to alteration.

The term "conservative substitution" is intended to mean that a substitution in which the amino acid residue is replaced with an amino acid residue having a similar side chain. These families are known in the art and include amino acids with basic side chains (e.g. lysine, arginine and histidine), acidic side chains (e.g. aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagines, glutamine, serine, threonine, tyrosine, cysteine), non-polar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine).

Functional nucleic acid equivalents may typically contain silent mutations or mutations that do not alter the biological function of encoded polypeptide. Accordingly, the invention provides nucleic acid molecules encoding lipolytic enzyme proteins that contain changes in amino acid residues that are not essential for a particular biological activity. Such lipolytic enzyme proteins differ in amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 yet retain at least one biological activity. In one embodiment the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises a substantially homologous amino acid sequence of at least about 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39.

For example, guidance concerning how to make phenotypically silent amino acid substitutions is provided in Bowie, J.U. et al., Science 247:1306-1310 (1990) wherein the authors indicate that there are two main approaches for studying the tolerance of an amino acid sequence to change. The first method relies on the process of evolution, in which mutations are either accepted or rejected by natural selection. The

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second approach uses genetic engineering to introduce amino acid changes at specific positions of a cloned gene and selects or screens to identify sequences that maintain functionality. As the authors state, these studies have revealed that proteins are surprisingly tolerant of amino acid substitutions. The authors further indicate which
5 changes are likely to be permissive at a certain position of the protein. For example, most buried amino acid residues require non-polar side chains, whereas few features of surface side chains are generally conserved. Other such phenotypically silent substitutions are described in Bowie et al, supra, and the references cited therein.

An isolated nucleic acid molecule encoding a protein homologous to the protein
10 selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 can be created by introducing one or more nucleotide substitutions, additions or deletions into the corresponding coding nucleotide sequences (Table 1) such that one or more amino acid substitutions, deletions or insertions are introduced into the encoded
15 protein. Such mutations may be introduced by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis.

The term "functional equivalents" also encompasses orthologues of the *Aspergillus niger* lipolytic enzymes provided herein. Orthologues of the *Aspergillus niger* lipolytic enzymes are proteins that can be isolated from other strains or species and possess a similar or identical biological activity. Such orthologues can readily be
20 identified as comprising an amino acid sequence that is substantially homologous to the amino acid sequences selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39.

As defined herein, the term "substantially homologous" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical
25 or equivalent (e.g., with similar side chain) amino acids or nucleotides to a second amino acid or nucleotide sequence such that the first and the second amino acid or nucleotide sequences have a common domain. For example, amino acid or nucleotide sequences which contain a common domain having about 60%, preferably 65%, more preferably 70%, even more preferably 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identity
30 or more are defined herein as sufficiently identical.

Also, nucleic acids encoding other lipolytic enzyme family members, which thus have a nucleotide sequence that differs from a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25,

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26, 28, 29, 31, 32, 34, 35, 37 and 38, are within the scope of the invention. Moreover, nucleic acids encoding lipolytic enzyme proteins from different species which thus have a nucleotide sequence which differs from a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 5 28, 29, 31, 32, 34, 35, 37 and 38 are within the scope of the invention.

Nucleic acid molecules corresponding to variants (e.g. natural allelic variants) and homologues of the polynucleotides of the invention can be isolated based on their homology to the nucleic acids disclosed herein using the cDNAs disclosed herein or a suitable fragment thereof, as a hybridisation probe according to standard hybridisation 10 techniques preferably under highly stringent hybridisation conditions.

In addition to naturally occurring allelic variants of the *Aspergillus niger* sequences provided herein, the skilled person will recognise that changes can be introduced by mutation into the nucleotide sequences selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 15 32, 34, 35, 37 and 38 thereby leading to changes in the amino acid sequence of the lipolytic enzyme protein without substantially altering the function of the protein.

In another aspect of the invention, improved lipolytic enzymes are provided. Improved lipolytic enzymes are proteins wherein at least one biological activity is improved. Such proteins may be obtained by randomly introducing mutations along all or 20 part of the lipolytic enzyme coding sequence, such as by saturation mutagenesis, and the resulting mutants can be expressed recombinantly and screened for biological activity. For instance, the art provides for standard assays for measuring the enzymatic activity of lipolytic enzymes and thus improved proteins may easily be selected.

In a preferred embodiment the lipolytic enzyme has an amino acid sequence 25 selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39. In another embodiment, the lipolytic enzyme is substantially homologous to the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 and retains at least one biological activity of a polypeptide selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 30 27, 30, 33, 36 and 39, yet differs in amino acid sequence due to natural variation or mutagenesis as described above.

In a further preferred embodiment, the lipolytic enzyme has an amino acid sequence encoded by an isolated nucleic acid fragment capable of hybridising to a

nucleic acid selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26, 28, 29, 31, 32, 34, 35, 37 and 38, preferably under highly stringent hybridisation conditions.

Accordingly, the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 and retains at least one functional activity of the polypeptide selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39.

10 In particular, the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 3 or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 6, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 9, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 12 or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 15, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 18 or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 21, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 24 or the lipolytic enzyme is a protein which comprises an amino

acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 27, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 30 or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 33, or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 36 or the lipolytic enzyme is a protein which comprises an amino acid sequence at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more homologous to the amino acid sequence shown in SEQ ID NO: 39.

Functional equivalents of a protein according to the invention can also be identified e.g. by screening combinatorial libraries of mutants, e.g. truncation mutants, of the protein of the invention for lipolytic enzyme activity. In one embodiment, a variegated library of variants is generated by combinatorial mutagenesis at the nucleic acid level. A variegated library of variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential protein sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage display). There are a variety of methods that can be used to produce libraries of potential variants of the polypeptides of the invention from a degenerate oligonucleotide sequence. Methods for synthesizing degenerate oligonucleotides are known in the art (see, e.g., Narang (1983) Tetrahedron 39:3; Itakura et al. (1984) Annu. Rev. Biochem. 53:323; Itakura et al. (1984) Science 198:1056; Ike et al. (1983) Nucleic Acid Res. 11:477).

In addition, libraries of fragments of the coding sequence of a polypeptide of the invention can be used to generate a variegated population of polypeptides for screening a subsequent selection of variants. For example, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of the coding sequence of interest with a nuclease under conditions wherein nicking occurs only about once per

molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal and internal fragments of various sizes of the protein of interest.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations of truncation, and for screening cDNA libraries for gene products having a selected property. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recursive ensemble mutagenesis (REM), a technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify variants of a protein of the invention (Arkin and Yourvan (1992) Proc. Natl. Acad. Sci. USA 89:7811-7815; Delgrave et al. (1993) Protein Engineering 6(3):327-331).

It will be apparent for the person skilled in the art that DNA sequence polymorphisms that may lead to changes in the amino acid sequence of the lipolytic enzyme may exist within a given population. Such genetic polymorphisms may exist in cells from different populations or within a population due to natural allelic variation. Allelic variants may also include functional equivalents.

Fragments of a polynucleotide according to the invention may also comprise polynucleotides not encoding functional polypeptides. Such polynucleotides may function as probes or primers for a PCR reaction.

Nucleic acids according to the invention irrespective of whether they encode functional or non-functional polypeptides, can be used as hybridization probes or polymerase chain reaction (PCR) primers. Uses of the nucleic acid molecules of the present invention that do not encode a polypeptide having a lipolytic enzyme activity include, inter alia, (1) isolating the gene encoding the lipolytic enzyme protein, or allelic variants thereof from a cDNA library e.g. from other organisms than *Aspergillus niger*; (2) in situ hybridization (e.g. FISH) to metaphase chromosomal spreads to provide precise

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chromosomal location of the lipolytic enzyme gene as described in Verma et al., Human Chromosomes: a Manual of Basic Techniques, Pergamon Press, New York (1988); (3) Northern blot analysis for detecting expression of lipolytic enzyme mRNA in specific tissues and/or cells and 4) probes and primers that can be used as a diagnostic tool to
5 analyse the presence of a nucleic acid hybridisable to the lipolytic enzyme probe in a given biological (e.g. tissue) sample.

Also encompassed by the invention is a method of obtaining a functional equivalent of a lipolytic enzyme-encoding gene or cDNA. Such a method entails obtaining a labelled probe that includes an isolated nucleic acid which encodes all or a
10 portion of the sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or a variant thereof; screening a nucleic acid fragment library with the labelled probe under conditions that allow hybridisation of the probe to nucleic acid fragments in the library, thereby forming nucleic acid duplexes, and preparing a full-length gene sequence from the nucleic acid fragments in any labelled
15 duplex to obtain a gene related to the lipolytic enzyme gene.

In one embodiment, a nucleic acid of the invention is at least 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more homologous to a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 2, 4, 5, 7, 8, 10, 11, 13, 14, 16, 17, 19, 20, 22, 23, 25, 26,
20 28, 29, 31, 32, 34, 35, 37 and 38 or the complement thereof.

In another preferred embodiment a polypeptide of the invention is at least 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more homologous to an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39.

25

Host cells

In another embodiment, the invention features cells, e.g., transformed host cells or recombinant host cells that contain a nucleic acid encompassed by the invention. A
30 "transformed cell" or "recombinant cell" is a cell into which (or into an ancestor of which) has been introduced, by means of recombinant DNA techniques, a nucleic acid according to the invention. Both prokaryotic and eukaryotic cells are included, e.g., bacteria, fungi, yeast, and the like, especially preferred are cells from filamentous fungi,

in particular *Aspergillus niger*.

A host cell can be chosen that modulates the expression of the inserted sequences, or modifies and processes the gene product in a specific, desired fashion. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may facilitate optimal functioning of the protein.

Various host cells have characteristic and specific mechanisms for post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems familiar to those of skill in the art of molecular biology and/or microbiology can be chosen to ensure the desired and correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells that possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product can be used. Such host cells are well known in the art.

Host cells also include, but are not limited to, mammalian cell lines such as CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, and choroid plexus cell lines.

If desired, the polypeptides according to the invention can be produced by a stably-transfected cell line. A number of vectors suitable for stable transfection of mammalian cells are available to the public, methods for constructing such cell lines are also publicly known, e.g., in Ausubel et al. (supra).

20

Antibodies

The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind lipolytic enzyme proteins according to the invention.

As used herein, the term "antibody" (Ab) or "monoclonal antibody" (Mab) is meant to include intact molecules as well as antibody fragments (such as, for example, Fab and F(ab')₂ fragments) which are capable of specifically binding to lipolytic enzyme protein. Fab and F(ab')₂ fragments lack the Fc fragment of intact antibody, clear more rapidly from the circulation, and may have less non-specific tissue binding of an intact antibody (Wahl et al., *J. Nucl. Med.* 24:316-325 (1983)). Thus, these fragments are preferred.

The antibodies of the present invention may be prepared by any of a variety of methods. For example, cells expressing the lipolytic enzyme protein or an antigenic

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fragment thereof can be administered to an animal in order to induce the production of sera containing polyclonal antibodies. In a preferred method, a preparation of lipolytic enzyme protein is prepared and purified to render it substantially free of natural contaminants. Such a preparation is then introduced into an animal in order to produce
5 polyclonal antisera of greater specific activity.

In the most preferred method, the antibodies of the present invention are monoclonal antibodies (or lipolytic enzyme protein binding fragments thereof). Such monoclonal antibodies can be prepared using hybridoma technology (Kohler *et al.*, *Nature* 256:495 (1975); Kohler *et al.*, *Eur. J. Immunol.* 6:511 (1976); Hammerling *et al.*,
10 *In: Monoclonal Antibodies and T-Cell Hybridomas*, Elsevier, N.Y., pp. 563-681 (1981)). In general, such procedures involve immunizing an animal (preferably a mouse) with a lipolytic enzyme protein antigen or, with a lipolytic enzyme protein expressing cell. The splenocytes of such mice are extracted and fused with a suitable myeloma cell line. Any suitable myeloma cell line may be employed in accordance with the present invention;
15 however, it is preferably to employ the parent myeloma cell line (SP₂O), available from the American Type Culture Collection, Rockville, Maryland. After fusion, the resulting hybridoma cells are selectively maintained in HAT medium, and then cloned by limiting dilution as described by Wands *et al.* (*Gastro-enterology* 80:225-232 (1981)). The hybridoma cells obtained through such a selection are then assayed to identify clones
20 which secrete antibodies capable of binding the lipolytic enzyme protein antigen. In general, the polypeptides can be coupled to a carrier protein, such as KLH, as described in Ausubel *et al.*, *supra*, mixed with an adjuvant, and injected into a host mammal.

In particular, various host animals can be immunized by injection of a polypeptide of interest. Examples of suitable host animals include rabbits, mice, guinea
25 pigs, and rats. Various adjuvants can be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), adjuvant mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, BCG (bacille Calmette-Guerin) and
30 *Corynebacterium parvum*. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

Such antibodies can be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD, and any subclass thereof. The hybridomas producing the mAbs of this

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invention can be cultivated *in vitro* or *in vivo*.

Once produced, polyclonal or monoclonal antibodies are tested for specific recognition of a protein according to the invention or functional equivalent thereof in an immunoassay, such as a Western blot or immunoprecipitation analysis using standard techniques, e.g., as described in Ausubel et al., *supra*. Antibodies that specifically bind to a protein according to the invention or functional equivalents thereof are useful in the invention. For example, such antibodies can be used in an immunoassay to detect a protein according to the invention in pathogenic or non-pathogenic strains of *Aspergillus* (e.g., in *Aspergillus* extracts).

Preferably, antibodies of the invention are produced using fragments of a protein according to the invention that appear likely to be antigenic, by criteria such as high frequency of charged residues. For example, such fragments may be generated by standard techniques of PCR, and then cloned into the pGEX expression vector (Ausubel et al., *supra*). Fusion proteins may then be expressed in *E. coli* and purified using a glutathione agarose affinity matrix as described in Ausubel, et al., *supra*. If desired, several (e.g., two or three) fusions can be generated for each protein, and each fusion can be injected into at least two rabbits. Antisera can be raised by injections in a series, typically including at least three booster injections. Typically, the antisera are checked for their ability to immunoprecipitate the protein according to the invention or functional equivalents thereof whereas unrelated proteins may serve as a control for the specificity of the immune reaction.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778 and 4,704,692) can be adapted to produce single chain antibodies against a protein according to the invention or functional equivalents thereof. Kits for generating and screening phage display libraries are commercially available e.g. from Pharmacia.

Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. Patent No. 5,223, 409; PCT Publication No. WO 92/18619; PCT Publication No. WO 91/17271; PCT Publication No. WO 20791; PCT Publication No. WO 92/20791; PCT Publication No. WO 92/15679; PCT Publication No. WO 93/01288; PCT Publication No. WO 92/01047; PCT Publication No. WO 92/09690; PCT Publication No. WO 90/02809; Fuchs et al. (1991) *BioTechnology* 9:1370-1372; Hay et al. (1992) *Hum. Antibod.*

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Hybridomas 3:81-85; Huse et al. (1989) *Science* 246:1275-1281; Griffiths et al. (1993) *EMBO J.* 12:725-734.

Polyclonal and monoclonal antibodies that specifically bind a protein according to the invention of functional equivalents thereof can be used, for example, to detect
5 expression of a gene encoding a protein according to the invention or a functional equivalent thereof e.g. in another strain of *Aspergillus*. For example, a protein according to the invention can be readily detected in conventional immunoassays of *Aspergillus* cells or extracts. Examples of suitable assays include, without limitation, Western blotting, ELISAs, radioimmune assays, and the like.

10 By "specifically binds" is meant that an antibody recognizes and binds a particular antigen, e.g., a protein according to the invention, but does not substantially recognize and bind other unrelated molecules in a sample.

Antibodies can be purified, for example, by affinity chromatography methods in which the polypeptide antigen is immobilized on a resin.

15 An antibody directed against a polypeptide of the invention (e.g., monoclonal antibody) can be used to isolate the polypeptide by standard techniques, such as affinity chromatography or immunoprecipitation. Moreover, such an antibody can be used to detect the protein (e.g., in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the polypeptide. The antibodies can also be
20 used diagnostically to monitor protein levels in cells or tissue as part of a clinical testing procedure, e.g., to, for example, determine the efficacy of a given treatment regimen or in the diagnosis of Aspergillosis..

Detection can be facilitated by coupling the antibody to a detectable substance.

25 Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β -galactosidase, or acetylcholinesterase; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a
30 luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive materials include ^{125}I , ^{131}I , ^{35}S or ^3H .

Preferred epitopes encompassed by the antigenic peptide are regions that are

located on the surface of the protein, e.g., hydrophilic regions. Hydrophobicity plots of the proteins of the invention can be used to identify hydrophilic regions.

The antigenic peptide of a protein of the invention comprises at least 7 (preferably 10, 15, 20, or 30) contiguous amino acid residues of the amino acid sequence selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 and encompasses an epitope of the protein such that an antibody raised against the peptide forms a specific immune complex with the protein.

Preferred epitopes encompassed by the antigenic peptide are regions of the protein according to the invention that are located on the surface of the protein, e.g., hydrophilic regions, hydrophobic regions, alpha regions, beta regions, coil regions, turn regions and flexible regions.

Immunoassays

Qualitative or quantitative determination of a polypeptide according to the present invention in a biological sample can occur using any art-known method. Antibody-based techniques provide special advantages for assaying specific polypeptide levels in a biological sample.

In these, the specific recognition is provided by the primary antibody (polyclonal or monoclonal) but the secondary detection system can utilize fluorescent, enzyme, or other conjugated secondary antibodies. As a result, an immunocomplex is obtained.

Accordingly, the invention provides a method for diagnosing whether a certain organism is infected with *Aspergillus* comprising the steps of:

- Isolating a biological sample from said organism suspected to be infected with *Aspergillus*,
- reacting said biological sample with an antibody according to the invention,
- determining whether immunocomplexes are formed.

Tissues can also be extracted, e.g., with urea and neutral detergent, for the liberation of protein for Western-blot or dot/slot assay. This technique can also be applied to body fluids.

Other antibody-based methods useful for detecting a protein according to the invention include immunoassays, such as the enzyme linked immunosorbent assay (ELISA) and the radioimmunoassay (RIA). For example, monoclonal antibodies against

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a protein according to the invention can be used both as an immunoabsorbent and as an enzyme-labeled probe to detect and quantify the protein according to the invention. The amount of protein present in the sample can be calculated by reference to the amount present in a standard preparation using a linear regression computer algorithm. In
5 another ELISA assay, two distinct specific monoclonal antibodies can be used to detect a protein according to the invention in a biological fluid. In this assay, one of the antibodies is used as the immuno-absorbent and the other as the enzyme-labeled probe.

The above techniques may be conducted essentially as a "one-step" or "two-step" assay. The "one-step" assay involves contacting a protein according to the
10 invention with immobilized antibody and, without washing, contacting the mixture with the labeled antibody. The "two-step" assay involves washing before contacting the mixture with the labeled antibody. Other conventional methods may also be employed as suitable. It is usually desirable to immobilize one component of the assay system on a support, thereby allowing other components of the system to be brought into contact with
15 the component and readily removed from the sample.

Suitable enzyme labels include, for example, those from the oxidase group, which catalyze the production of hydrogen peroxide by reacting with substrate. Activity of an oxidase label may be assayed by measuring the concentration of hydrogen peroxide formed by the enzyme-labelled antibody/substrate reaction.

20 Besides enzymes, other suitable labels include radioisotopes, such as iodine (^{125}I , ^{127}I), carbon (^{14}C), sulphur (^{35}S), tritium (^3H), indium (^{112}In), and technetium ($^{99\text{m}}\text{Tc}$), and fluorescent labels, such as fluorescein and rhodamine, and biotin.

Specific binding of a test compound to a protein according to the invention can be detected, for example, in vitro by reversibly or irreversibly immobilizing the protein
25 according to the invention on a substrate, e.g., the surface of a well of a 96-well polystyrene microtitre plate. Methods for immobilizing polypeptides and other small molecules are well known in the art. For example, the microtitre plates can be coated with a protein according to the invention by adding the protein in a solution (typically, at a concentration of 0.05 to 1 mg/ml in a volume of 1-100 μl) to each well, and incubating
30 the plates at room temperature to 37 $^{\circ}\text{C}$ for 0.1 to 36 hours. Proteins that are not bound to the plate can be removed by shaking the excess solution from the plate, and then washing the plate (once or repeatedly) with water or a buffer. Typically, the protein is contained in water or a buffer. The plate is then washed with a buffer that lacks the

bound protein. To block the free protein-binding sites on the plates, the plates are blocked with a protein that is unrelated to the bound protein. For example, 300 ul of bovine serum albumin (BSA) at a concentration of 2 mg/ml in Tris-HCl is suitable. Suitable substrates include those substrates that contain a defined cross-linking chemistry (e.g., plastic substrates, such as polystyrene, styrene, or polypropylene substrates from Corning Costar Corp. (Cambridge, MA), for example). If desired, a beaded particle, e.g., beaded agarose or beaded sepharose, can be used as the substrate.

Binding of the test compound to the polypeptides according to the invention can be detected by any of a variety of artknown methods. For example, a specific antibody can be used in an immunoassay. If desired, the antibody can be labeled (e.g., fluorescently or with a radioisotope) and detected directly (see, e.g., West and McMahon, J. Cell Biol. 74:264, 1977). Alternatively, a second antibody can be used for detection (e.g., a labeled antibody that binds the Fc portion of an anti-AN97 antibody). In an alternative detection method, the protein according to the invention is labeled, and the label is detected (e.g., by labeling a protein according to the invention with a radioisotope, fluorophore, chromophore, or the like). In still another method, the protein according to the invention is produced as a fusion protein with a protein that can be detected optically, e.g., green fluorescent protein (which can be detected under UV light). In an alternative method, the protein according to the invention can be covalently attached to or fused with an enzyme having a detectable enzymatic activity, such as horse radish peroxidase, alkaline phosphatase, alpha-galactosidase, or glucose oxidase. Genes encoding all of these enzymes have been cloned and are readily available for use by those of skill in the art. If desired, the fusion protein can include an antigen, and such an antigen can be detected and measured with a polyclonal or monoclonal antibody using conventional methods. Suitable antigens include enzymes (e.g., horse radish peroxidase, alkaline phosphatase, and alpha-galactosidase) and non-enzymatic polypeptides (e.g., serum proteins, such as BSA and globulins, and milk proteins, such as caseins).

Epitopes, antigens and immunogens.

In another aspect, the invention provides a peptide or polypeptide comprising

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an epitope-bearing portion of a polypeptide of the invention. The epitope of this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide of the invention. An "immunogenic epitope" is defined as a part of a protein that elicits an antibody response when the whole protein is the immunogen. These immunogenic epitopes are believed to be confined to a few loci on the molecule. On the other hand, a region of a protein molecule to which an antibody can bind is defined as an "antigenic epitope." The number of immunogenic epitopes of a protein generally is less than the number of antigenic epitopes. See, for instance, Geysen, H. M. et al., Proc. Natl. Acad. Sci. USA 81:3998-4002 (1984).

As to the selection of peptides or polypeptides bearing an antigenic epitope (i.e., that contain a region of a protein molecule to which an antibody can bind), it is well known in that art that relatively short synthetic peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See, for instance, Sutcliffe, J. G. et al., Science 219:660-666 (1984). Peptides capable of eliciting protein-reactive sera are frequently represented in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined neither to immunodominant regions of intact proteins (i.e., immunogenic epitopes) nor to the amino or carboxyl terminals. Peptides that are extremely hydrophobic and those of six or fewer residues generally are ineffective at inducing antibodies that bind to the mimicked protein; longer, soluble peptides, especially those containing proline residues, usually are effective. Sutcliffe et al., supra, at 661. For instance, 18 of 20 peptides designed according to these guidelines, containing 8-39 residues covering 75% of the sequence of the Influenza virus hemagglutinin HA1 polypeptide chain, induced antibodies that reacted with the HA1 protein or intact virus; and 12/12 peptides from the MuLV polymerase and 18/18 from the rabies glycoprotein induced antibodies that precipitated the respective proteins.

Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. Thus, a high proportion of hybridomas obtained by fusion of spleen cells from donors immunized with an antigen epitope-bearing peptide generally secrete antibody reactive with the native protein. Sutcliffe et al., supra, at 663. The antibodies raised by antigenic epitope bearing peptides or polypeptides are useful to detect the mimicked protein, and antibodies to different

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peptides may be used for tracking the fate of various regions of a protein precursor which undergoes posttranslation processing. The peptides and anti-peptide antibodies may be used in a variety of qualitative or quantitative assays for the mimicked protein, for instance in competition assays since it has been shown that even short peptides
5 (e.g., about 9 amino acids) can bind and displace the larger peptides in immunoprecipitation assays. See, for instance, Wilson, I.A. et al., Cell 37:767-778 at 777 (1984). The anti-peptide antibodies of the invention also are useful for purification of the mimicked protein, for instance, by adsorption chromatography using methods well known in the art.

10 Antigenic epitope-bearing peptides and polypeptides of the invention designed according to the above guidelines preferably contain a sequence of at least seven, more preferably at least nine and most preferably between about 15 to about 30 amino acids contained within the amino acid sequence of a polypeptide of the invention. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of a
15 polypeptide of the invention, containing about 30 to about 50 amino acids, or any length up to and including the entire amino acid sequence of a polypeptide of the invention, also are considered epitope-bearing peptides or polypeptides of the invention and also are useful for inducing antibodies that react with the mimicked protein. Preferably, the amino acid sequence of the epitope-bearing peptide is selected to provide substantial
20 solubility in aqueous solvents (i.e., the sequence includes relatively hydrophilic residues and highly hydrophobic sequences are preferably avoided); and sequences containing proline residues are particularly preferred.

The epitope-bearing peptides and polypeptides of the invention may be produced by any conventional means for making peptides or polypeptides including
25 recombinant means using nucleic acid molecules of the invention. For instance, a short epitope-bearing amino acid sequence may be fused to a larger polypeptide which acts as a carrier during recombinant production and purification, as well as during immunization to produce anti-peptide antibodies.

30 Epitope-bearing peptides also may be synthesized using known methods of chemical synthesis. For instance, Houghten has described a simple method for synthesis of large numbers of peptides, such as 10-20 mg of 248 different 13 residue peptides representing single amino acid variants of a segment of the HAI polypeptide which were prepared and characterized (by ELISA-type binding studies) in less than four

weeks. Houghten, R. A., Proc. Natl. Acad. Sci. USA 82:5131-5135 (1985). This "Simultaneous Multiple Peptide Synthesis (SMPS)" process is further described in U.S. Patent No. 4,631,211 to Houghten et al. (1986). In this procedure the individual resins for the solid-phase synthesis of various peptides are contained in separate solvent-permeable packets, enabling the optimal use of the many identical repetitive steps involved in solid-phase methods.

A completely manual procedure allows 500-1000 or more syntheses to be conducted simultaneously. Houghten et al., supra, at 5134.

Epitope-bearing peptides and polypeptides of the invention are used to induce antibodies according to methods well known in the art. See, for instance, Sutcliffe et al., supra; Wilson et al., supra; Chow, M. et al., Proc. Natl. Acad. Sci. USA 82:910-914; and Bittle, F.J. et al., J. Gen. Virol. 66:2347-2354 (1985).

Generally, animals may be immunized with free peptide; however, anti-peptide antibody titer may be boosted by coupling of the peptide to a macromolecular carrier, such as keyhole limpet hemocyanin (KLH) or tetanus toxoid. For instance, peptides containing cysteine may be coupled to carrier using a linker such as maleimidobenzoyl-N-hydroxysuccinimide ester (MBS), while other peptides may be coupled to carrier using a more general linking agent such as glutaraldehyde.

Animals such as rabbits, rats and mice are immunized with either free or carrier-coupled peptides, for instance, by intraperitoneal and/or intradermal injection of emulsions containing about 100 ug peptide or carrier protein and Freund's adjuvant. Several booster injections may be needed, for instance, at intervals of about two weeks, to provide a useful titer of anti-peptide antibody which can be detected, for example, by ELISA assay using free peptide adsorbed to a solid surface. The titer of anti-peptide antibodies in serum from an immunized animal may be increased by selection of anti-peptide antibodies, for instance, by adsorption to the peptide on a solid support and elution of the selected antibodies according to methods well known in the art.

Immunogenic epitope-bearing peptides of the invention, i.e., those parts of a protein that elicit an antibody response when the whole protein is the immunogen, are identified according to methods known in the art. For instance, Geysen et al., 1984, supra, discloses a procedure for rapid concurrent synthesis on solid supports of hundreds of peptides of sufficient purity to react in an enzyme-linked immunosorbent assay. Interaction of synthesized peptides with antibodies is then easily detected without

removing them from the support. In this manner a peptide bearing an immunogenic epitope of a desired protein may be identified routinely by one of ordinary skill in the art. For instance, the immunologically important epitope in the coat protein of foot-and-mouth disease virus was located by Geysen et al. with a resolution of seven amino acids by synthesis of an overlapping set of all 208 possible hexapeptides covering the entire 213 amino acid sequence of the protein. Then, a complete replacement set of peptides in which all 20 amino acids were substituted in turn at every position within the epitope were synthesized, and the particular amino acids conferring specificity for the reaction with antibody were determined. Thus, peptide analogs of the epitope-bearing peptides of the invention can be made routinely by this method. U.S. Patent No. 4,708,781 to Geysen (1987) further describes this method of identifying a peptide bearing an immunogenic epitope of a desired protein.

Further still, U.S. Patent No. 5,194,392 to Geysen (1990) describes a general method of detecting or determining the sequence of monomers (amino acids or other compounds) which is a topological equivalent of the epitope (i.e., a "mimotope") which is complementary to a particular paratope (antigen binding site) of an antibody of interest. More generally, U.S. Patent No. 4,433,092 to Geysen (1989) describes a method of detecting or determining a sequence of monomers which is a topographical equivalent of a ligand which is complementary to the ligand binding site of a particular receptor of interest. Similarly, U.S. Patent No. 5,480,971 to Houghten, R. A. et al. (1996) on Peralkylated Oligopeptide Mixtures discloses linear C1-C7-alkyl peralkylated oligopeptides and sets and libraries of such peptides, as well as methods for using such oligopeptide sets and libraries for determining the sequence of a peralkylated oligopeptide that preferentially binds to an acceptor molecule of interest. Thus, non-peptide analogs of the epitope-bearing peptides of the invention also can be made routinely by these methods.

Use of lipolytic enzymes in industrial processes

The invention also relates to the use of the lipolytic enzyme according to the invention in a selected number of industrial processes. Despite the long-term experience obtained with these processes, the lipolytic enzyme according to the invention features a number of significant advantages over the enzymes currently used. Depending on the

specific application, these advantages can include aspects like lower production costs, higher specificity towards the substrate, less antigenic, less undesirable side activities, higher yields when produced in a suitable microorganism, more suitable pH and temperature ranges, better tastes of the final product as well as food grade and kosher aspects.

5 The present invention also relates to methods for preparing a dough or a baked product comprising incorporating into the dough an effective amount of a lipolytic enzyme of the present invention which improves one or more properties of the dough or the baked product obtained from the dough relative to a dough or a baked product in
10 which the polypeptide is not incorporated.

The phrase "incorporating into the dough" is defined herein as adding the lipolytic enzyme according to the invention to the dough, any ingredient from which the dough is to be made, and/or any mixture of dough ingredients from which the dough is to be made. In other words, the lipolytic enzyme according to the invention may be added
15 in any step of the dough preparation and may be added in one, two or more steps. The lipolytic enzyme according to the invention is added to the ingredients of a dough that is kneaded and baked to make the baked product using methods well known in the art. See, for example, U.S. Patent No. 4,567,046, EP-A-426,211, JP-A-60-78529, JP-A-62-111629, and JP-A-63-258528.

20 The term "effective amount" is defined herein as an amount of the lipolytic enzyme according to the invention that is sufficient for providing a measurable effect on at least one property of interest of the dough and/or baked product.

The term "improved property" is defined herein as any property of a dough and/or a product obtained from the dough, particularly a baked product, which is
25 improved by the action of the lipolytic enzyme according to the invention relative to a dough or product in which the lipolytic enzyme according to the invention is not incorporated. The improved property may include, but is not limited to, increased strength of the dough, increased elasticity of the dough, increased stability of the dough, reduced stickiness of the dough, improved extensibility of the dough, improved flavour of
30 the baked product, improved anti-staling of the baked product.

The improved property may be determined by comparison of a dough and/or a baked product prepared with and without addition of a polypeptide of the present invention in accordance with the methods of present invention are described below in

the Examples. Organoleptic qualities may be evaluated using procedures well established in the baking industry, and may include, for example, the use of a panel of trained taste-testers.

5 The term "increased strength of the dough" is defined herein as the property of a dough that has generally more elastic properties and/or requires more work input to mould and shape.

The term "increased elasticity of the dough" is defined herein as the property of a dough which has a higher tendency to regain its original shape after being subjected to a certain physical strain.

10 The term "increased stability of the dough" is defined herein as the property of a dough that is less susceptible to mechanical abuse thus better maintaining its shape and volume.

The term "reduced stickiness of the dough" is defined herein as the property of a dough that has less tendency to adhere to surfaces, e.g., in the dough production machinery, and is either evaluated empirically by the skilled test baker or measured by the use of a texture analyser (e.g., TAXT2) as known in the art.

The term "improved extensibility of the dough" is defined herein as the property of a dough that can be subjected to increased strain or stretching without rupture.

20 The term "improved machineability of the dough" is defined herein as the property of a dough that is generally less sticky and/or more firm and/or more elastic.

The term "increased volume of the baked product" is measured as the specific volume of a given loaf of bread (volume/weight) determined typically by the traditional rapeseed displacement method.

25 The term "improved crumb structure of the baked product" is defined herein as the property of a baked product with finer and/or thinner cell walls in the crumb and/or more uniform/homogenous distribution of cells in the crumb and is usually evaluated empirically by the skilled test baker.

30 The term "improved softness of the baked product" is the opposite of "firmness" and is defined herein as the property of a baked product that is more easily compressed and is evaluated either empirically by the skilled test baker or measured by the use of a texture analyzer (e.g., TAXT2) as known in the art.

The term "improved flavor of the baked product" is evaluated by a trained test panel.

The term "improved anti-staling of the baked product" is defined herein as the properties of a baked product that have a reduced rate of deterioration of quality parameters, e.g., softness and/or elasticity, during storage.

5 The term "dough" is defined herein as a mixture of flour and other ingredients firm enough to knead or roll. The dough may be fresh, frozen, pre-bared, or pre-baked. The preparation of frozen dough is described by Kulp and Lorenz in Frozen and Refrigerated Doughs and Batters.

10 The term "baked product" is defined herein as any product prepared from a dough, either of a soft or a crisp character. Examples of baked products, whether of a white, light or dark type, which may be advantageously produced by the present invention are bread (in particular white, whole-meal or rye bread), typically in the form of loaves or rolls, French baguette-type bread, pasta, pita bread, tortillas, tacos, cakes, pancakes, biscuits, cookies, pie crusts, steamed bread, and crisp bread, and the like.

15 Lipolytic enzyme of the present invention and/or additional enzymes to be used in the methods of the present invention may be in any form suitable for the use in question, e.g., in the form of a dry powder, agglomerated powder, or granulate, in particular a non-dusting granulate, liquid, in particular a stabilized liquid, or protected enzyme such described in WO01/11974 and WO02/26044. Granulates and agglomerated powders may be prepared by conventional methods, e.g., by spraying the
20 lipolytic enzyme according to the invention onto a carrier in a fluid-bed granulator. The carrier may consist of particulate cores having a suitable particle size. The carrier may be soluble or insoluble, e.g., a salt (such as NaCl or sodium sulphate), sugar (such as sucrose or lactose), sugar alcohol (such as sorbitol), starch, rice, corn grits, or soy. The lipolytic enzyme according to the invention and/or additional enzymes may be contained
25 in slow-release formulations. Methods for preparing slow-release formulations are well known in the art. Adding nutritionally acceptable stabilizers such as sugar, sugar alcohol, or another polyol, and/or lactic acid or another organic acid according to established methods may for instance, stabilize liquid enzyme preparations.

30 The lipolytic enzyme according to the invention may also be incorporated in yeast comprising compositions such as disclosed in EP-A-0619947, EP-A-0659344 and WO02/49441.

For inclusion in pre-mixes of flour it is advantageous that the polypeptide according to the invention is in the form of a dry product, e.g., a non-dusting granulate,

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whereas for inclusion together with a liquid it is advantageously in a liquid form.

One or more additional enzymes may also be incorporated into the dough. The additional enzyme may be of any origin, including mammalian and plant, and preferably of microbial (bacterial, yeast or fungal) origin and may be obtained by techniques
5 conventionally used in the art.

In a preferred embodiment, the additional enzyme may be an amylase, such as an alpha-amylase (useful for providing sugars fermentable by yeast and retarding staling) or beta-amylase, cyclodextrin glucoamylase, peptidase, in particular, an
10 exopeptidase (useful in flavour enhancement), transglutaminase, lipase (useful for the modification of lipids present in the dough or dough constituents so as to soften the dough), phospholipase, cellulase, hemicellulase, in particular a pentosanase such as
15 xylanase (useful for the partial hydrolysis of pentosans which increases the extensibility of the dough), protease (useful for gluten weakening in particular when using hard wheat flour), protein disulfide isomerase, e.g., a protein disulfide isomerase as disclosed in WO
95/00636, glycosyltransferase, peroxidase (useful for improving the dough consistency),
laccase, or oxidase, e.g., an glucose oxidase, hexose oxidase, aldose oxidase, pyranose oxidase, lipoxygenase or L-amino acid oxidase (useful in improving dough consistency).

When one or more additional enzyme activities are to be added in accordance
20 with the methods of the present invention, these activities may be added separately or together with the polypeptide according to the invention, optionally as constituent(s) of the bread-improving and/or dough-improving composition. The other enzyme activities may be any of the enzymes described above and may be dosed in accordance with
established baking practices.

25 The present invention also relates to methods for preparing a baked product comprising baking a dough obtained by a method of the present invention to produce a baked product. The baking of the dough to produce a baked product may be performed using methods well known in the art.

The present invention also relates to doughs and baked products, respectively,
30 produced by the methods of the present invention.

The present invention further relates to a pre-mix, e.g., in the form of a flour composition, for dough and/or baked products made from dough, in which the pre-mix comprises a polypeptide of the present invention. The term "pre-mix" is defined herein to

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be understood in its conventional meaning, i.e., as a mix of baking agents, generally including flour, which may be used not only in industrial bread-baking plants/facilities, but also in retail bakeries. The pre-mix may be prepared by mixing the polypeptide or a bread-improving and/or dough-improving composition of the invention comprising the polypeptide with a suitable carrier such as flour, starch, a sugar, or a salt. The pre-mix may contain other dough-improving and/or bread-improving additives, e.g., any of the additives, including enzymes, mentioned above.

The present invention further relates to baking additives in the form of a granulate or agglomerated powder, which comprise a polypeptide of the present invention. The baking additive preferably has a narrow particle size distribution with more than 95% (by weight) of the particles in the range from 25 to 500 μm .

In dough and bread making the present invention may be used in combination with the processing aids defined hereinbefore such as the chemical processing aids like oxidants (e.g. ascorbic acid), reducing agents (e.g. L-cysteine), oxidoreductases (e.g. glucose oxidase) and/or other enzymes such as polysaccharide modifying enzymes (e.g. α -amylase, hemicellulase, branching enzymes, etc.) and/or protein modifying enzymes (endoprotease, exoprotease, branching enzymes, etc.).

EXAMPLE 1

20

Fermentation of *Aspergillus niger*

Lipolytic enzymes encoded by the nucleotide sequence as provided herein were obtained by constructing expression plasmids containing the DNA sequences, transforming an *A. niger* strain with this plasmid and growing the *Aspergillus niger* strains in the following way.

Fresh spores (10^6 - 10^7) of *A. niger* strains were inoculated in 20 ml CSL-medium (100 ml flask, baffle) and grown for 20-24 hours at 34°C and 170 rpm. After inoculation of 5-10 ml CSL pre-culture in 100 ml CSM medium (500 ml flask, baffle) the strains were fermented at 34°C and 170 rpm for 3-5 days.

Cell-free supernatants were obtained by centrifugation in 50 ml Greiner tubes (30 minutes, 5000 rpm). The supernatants were pre-filtered over a GF/A Whatman Glass microfiber filter (150 mm ϕ) to remove the larger particles, adjusted to pH 5 with 4 N

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KOH (if necessary) and sterile filtrated over a 0.2 µm (bottle-top) filter with suction to remove the fungal material. The supernatants were stored at 4°C (or -20°C).

The CSL medium consisted of (in amount per litre): 100 g Corn Steep Solids (Roquette), 1 g NaH₂PO₄·H₂O, 0.5 g MgSO₄·7H₂O, 10 g glucose·H₂O and 0.25 g
5 Basildon (antifoam). The ingredients were dissolved in demi-water and the pH was adjusted to pH 5.8 with NaOH or H₂SO₄; 100 ml flasks with baffle and foam ball were filled with 20 ml fermentation broth and sterilized for 20 minutes at 120°C after which 200 µl of a solution containing 5000 IU/ml penicillin and 5 mg/ml Streptomycin was added to each flask after cooling to room temperature.

10 The CSM medium consisted of (in amount per litre): 150 g maltose·H₂O, 60 g Soytone (pepton), 1 g NaH₂PO₄·H₂O, 15 g MgSO₄·7H₂O, 0.08 g Tween 80, 0.02 g Basildon (antifoam), 20 g MES, 1 g L-arginine. The ingredients were dissolved in demi-water and the pH was adjusted to pH 6.2 with NaOH or H₂SO₄; 500 ml flasks with baffle and foam ball were filled with 100 ml fermentation broth and sterilized for 20 minutes at
15 120°C after which 1 ml of a solution containing 5000 IU/ml penicillin and 5 mg/ml Streptomycin was added to each flask after cooling to room temperature.

EXAMPLE 2

Purification of the lipolytic enzymes of the invention

20

Step 1 - Preparation of ultrafiltrates

The supernatants of the cultures, as obtained in Example 1, were ultrafiltrated to remove the low molecular contaminations that could interfere with the enzymatic activity determinations and the baking tests. Ultrafiltration of 30 ml supernatant was performed in
25 a Millipore Labscale TFF system equipped with a filter with a 10 kDa cut-off.

Depending on their colour, the samples were washed 3-5 times with 40 ml volumes of cold 100 mM phosphate buffer pH 6.0 including 0.5 mM CaCl₂. The final volume of the enzyme solution was 30 ml and is further referred to as "ultrafiltrate".

30 *Step 2 - Determination of the lipolytic enzymes concentration by A280 and HPSEC.*

The concentration of the lipolytic enzymes in the ultrafiltrate was calculated from the extinction at 280 nm (A280) attributable to the lipolytic enzymes and the calculated molecular extinction coefficient of the lipolytic enzymes. Measurement of the A280 was

performed in an Uvikon XL Secomam spectrophotometer (Beun de Ronde, Abcoude, The Netherlands).

The molecular extinction coefficient of an enzyme can be calculated from the number of tyrosine, tryptophan and cysteine residues per enzyme molecule (S.C. Gill and P.H. von Hippel, Anal. Biochem. 182, 319-326 (1989)). The molecular extinction coefficient of these amino acids are 1280, 5690 and 120 M⁻¹.cm⁻¹ respectively. The number of tyrosine, tryptophan and cysteine residues in the lipolytic enzymes of the invention can be deduced from the protein sequences selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39. The calculated extinction coefficients of the lipolytic enzymes of the invention are summarized in Table 2.

Table 2

Lipolytic enzyme	SEQ ID NO:	# amino acids			Calculated M.W. (Da)	Calculated extinction coefficient at 280 nm	
		Trp	Tyr	Cys		M ⁻¹ .cm ⁻¹	(1 mg/ml) ⁻¹ .cm ⁻¹
NBE028	3	13	26	6	64141	107970	1.7
NBE029	6	14	27	6	63250	114940	1.8
NBE030	9	17	26	6	59952	130730	2.2
NBE031	12	9	27	4	61173	86250	1.4
NBE032	15	3	13	6	29683	34430	1.2
NBE033	18	7	24	2	44890	70790	1.6
NBE034	21	11	19	7	53796	87750	1.6
NBE036	24	10	23	7	64945	87180	1.3
NBE038	27	13	29	4	55161	111570	2.2
NBE039	30	11	26	6	59298	96590	1.6
NBE043	33	16	35	8	62564	136800	2.2
NBE045	36	0	6	6	26688	8400	0.31
NBE042	39	14	30	7	61593	118900	1.9

The extinction of the ultrafiltrate at 280 nm (A280) that is attributable to the lipolytic enzymes depends on the purity of the enzyme sample. This purity was

determined using HPSEC (High Performance Size Exclusion Chromatography) with a TSK SW-XL column (300*7,8 mm; MW range 10-300 kDa). The elution buffer consisted of 25 mM sodium phosphate buffer pH 6.0 and was used at a flow of 1 ml/min. Samples of 5—100 µl were injected. The absorbance at 280 nm was measured.

5 The A280 in the ultrafiltrate attributable to the lipolytic enzyme of the invention was obtained from the ratio of the peak surface of the respective lipolytic enzyme peak in the chromatogram and the total surface of the peaks absorbing at 280 nm. The lipolytic enzyme concentration in the ultrafiltrate was then calculated by multiplying the A280 of the ultrafiltrate by the ratio described above and divided by the calculated extinction
10 coefficient (1 mg/ml solution – Table 2 most right column) for each lipolytic enzyme.

EXAMPLE 3

Activity measurements

The cell-free supernatants obtained in Example 1 were subjected to the lipase,
15 phospholipase and galactolipase assays as summarized in Table 3.

Table 3. Lipolytic enzyme activities in the cell free supernatants as prepared in Example 1.

Lipolytic enzyme	Lipase	phospho lipase A	lyso phospho lipase	galacto lipase
NBE028	+	+	+	0
NBE029	+	+	+	0
NBE031	+++	+	+	+
NBE032	++	+	+	0
NBE033	+	++	+	+
NBE034	0	+	0	0
NBE036	0	+	+	0
NBE038	0	+	0	0
NBE039	+	0	0	0
NBE043	+	0	0	0

0 = not different from blanc; +/++/+++ = higher than blanc;

20 Lipase activity was determined spectrophotometrically by using 2,3-mercapto-1-propanol-tributyrate (TBDMP) as a substrate. Lipase hydrolyses the sulphide bond of

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TBDMP thereby liberating thio-butanoic acid which in a subsequent reaction with 4,4,-
dithiodipyridine (DTDP) forms 4-thiopyridone. The latter is in a tautomeric equilibrium
with 4-mercaptopyridine which absorbs at 334 nm. The reaction is carried out in 0.1 M
acetate buffer pH 5.0 containing 0.2 % Triton-X100, 0.65 mM TBDMP and 0.2 mM DTDP
5 at 37°C. One lipase unit is defined as the amount of enzyme that liberates 1 micromole
of 4 thio-butanoic acid per minute at the reaction conditions stated.

Phospholipase A was determined spectrophotometrically by using 1,2-
dithiooctanoyl-phosphatidylcholine as a substrate. Phospholipase A hydrolyses the
10 sulphide bond at the 1 position (PLA1) or the 2 position (PLA2) thereby liberating 4 thio-
octanoic acid which, in a subsequent reaction reacts with 4,4'-dithiopyridine to form 4-
thiopyridone. The latter is in tautomeric equilibrium with 4-mercaptopyridine that absorbs
at 334 nm. The reaction is carried out in 0.1 M acetate buffer pH 4.0 containing 0.2 %
Triton-X100, 0.65 mM substrate and 0.2 mM DTDP at 37°C. One phospholipase A unit
15 (PLA) is defined as the amount of enzyme that liberates 1 micromole of 4 thio-octanoic
acid per minute at the reaction conditions stated.

Lysophospholipase activity was determined with ³¹P-NMR spectroscopy by
using lysophosphatidyl-choline as a substrate. Lysophospholipase hydrolyses the ester
20 bond thereby liberating the fatty acid from the glycerol moiety. The so-formed
glycerolphosphocholine is quantified using NMR.

The reaction is carried out in 50 mM acetic acid buffer pH 4.5 further containing 1 mg/ml
lysophosphatidylcholine and 5 mM CaCl₂ for 30 minutes at 55°C.

One lysophospholipase unit (LPC) is defined as the amount of enzyme that forms 1
25 micromole of 4 glycerolphosphocholine per minute at the reaction conditions stated.

Galactolipase activity was determined with ¹H-NMR spectroscopy by using
digalactosyldiglyceride as a substrate, according to the method described by Hirayama
and Matsuda (1972) Agric. Biol. Chem. 36, 1831. Galactolipase hydrolyses the ester
30 bond between the fatty acids and the glycerol backbone thereby liberating one or both
fatty acids. The reaction is carried out in 50 mM acetic acid buffer pH 4.5 further
containing 4 mM CaCl₂, 0.2% Triton X-100 and 1 mg/ml digalactosyldiglyceride (Lipid
Products) for 30 minutes at 30°C. One galactolipase unit is defined as the amount of

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enzyme that forms 1 micromole of fatty acid per minute at the reaction conditions stated.

The ultrafiltrates obtained in Example 2, were subjected to the FAU enzyme activity measurement. The activity of the fungal alpha-amylase was measured using Phadebas Amylase test tablets (Pharmacia). Phadebas tablets contain a water insoluble starch substrate and a blue dye, bound by cross-linking to the substrate. The substrate is hydrolysed by fungal amylase, releasing dyed soluble maltodextrines that go into solution. A calibration curve was prepared with a solution containing a reference fungal alpha amylase activity. From the reference and unknown samples appropriate dilutions were prepared in 50 mM malic acid buffer pH 5.5. Samples of 5 ml were incubated with 30°C for 5 minutes, a Phadebas tablet was added and after 15 minutes the reaction was stopped by the addition of 1.0 ml 0.5 N sodium hydroxide. The mixtures were allowed to cool down to room temperature for 5 minutes after which 4.0 ml water was added, shaken by hand and after 15 minutes the samples were centrifuged at 4700 rpm for 10 minutes. The extinction of the top layers was measured at 620 nm. The OD 620 nm is a measure for fungal alpha amylase activity. One fungal amylase unit (FAU) is defined herein as the amount of enzyme that converts 1 gram of starch (100% dry matter) per hour into a product having a transmission at 620 nm after reaction with a iodine solution of known strength at the reaction conditions stated.

20

Table 4. FAU and protein in the ultrafiltrates as prepared in Example 2.

lipolytic enzyme	Protein (mg/ml) from the 280 nm analysis	fungal amylase (FAU/ml)
NBE028	2.3	4.5
NBE029	1.3	3.0
NBE030	0.4	2.6
NBE031	0.1	2.5
NBE032	1.0	0.3
NBE033	ND	0.3

NBE034	ND	2.7
NBE036	ND	3.4
NBE038	2.0	3.7
NBE039	2.2	0.6
NBE043	0.1	0.2
NBE045	ND	4.0
NBE042	1.6	1.5

In addition to the activities mentioned in Table 4, minor activities of glucoamylase was also present, however in such low amounts that these enzymes did not interfere in the baking experiments described in example 4.

5

EXAMPLE 4

Baking experiments 1 – pup loaves

Pup loaves were baked from 150 gram dough pieces obtained by mixing 200 g flour (Kolibri™/Ibis™ in a ratio of 80/20), 1,4 g dried baker's yeast (Fermipan®), 4 g salt, 3 g sugar, 10 mg ascorbic acid, 116 g water and 2 g fat. After mixing for 6 minutes and 15 seconds in a pin mixer, the dough was divided into pieces of 150 grams and proofed for 45 minutes at 30°C, punched, proofed for another 25 minutes, moulded and panned. Proofing took place at a relative humidity of 90-100%. After a final proof of 70 minutes at 30°C, the dough was baked for 20 minutes at 225°C.

15

The various effects (Tables 5 and 6) of the different lipolytic enzymes in the baking experiments were compared with a control containing the same amount of fungal amylase that was added otherwise by the dosage of the ultrafiltrate (for the fungal amylase activity in the ultrafiltrates see Table 4). This was necessary since the amounts of fungal amylase added with the lipolytic enzymes in particular affected the loaf volume, not the other parameters. The volume of the breads with the control amount of fungal amylase added was taken as 100%.

20

Table 5.

effect		Score				
		1	2	3	4	5
Dough	dough stickiness	too sticky	sticky	control bread	much better	excellent dry
	dough extensibility	Too short	Shorter than the control	control bread	good	too long
baked bread	crumb structure	poor	non-uniform	control bread	good	excellent
	crust colour	Nearly white	too light	control bread	excellent	too dark
	crumb colour	Far too yellow	too yellow	control bread	excellent	absolutely white
	staling	Far too firm	too firm	control bread	softer	excellent

5 Loaf volume was determined by the Bread Volume Measurer BVM-3 (RI Cards Instruments AB, Viken, Sweden). The principle of this measurement is based on the reflection of ultrasound measured by a sensor around a rotating bread. A measurement time was taken of 45 seconds.

Dough stickiness and extensibility were evaluated by a qualified baker using the scale depicted in Table 5. The average of 2 loaves per object was measured.

10 After these tests the dough pieces were rounded and a first proof was performed for 45 minutes at 30°C and hereafter the dough was punched, moulded, panned, proofed for 75 minutes at 30°C. The relative humidity during the proofs was set at 85%.

15 Subsequently the stability of the proofed dough was judged by the presence of bladders, torn side crust and irregular curved surfaces of the crust. The dough pieces were baked for 20 minutes at 225°C. Loaf volumes were determined by the BVM-3 method. In the table the average is presented of 2 breads that are baked from the same object.

The crumb structure was judged by a qualified baker using the scale depicted in Table 5. After storing the loaves for three days in polyethylene bags at room temperature crumb firmness was measured using a Stevens Texture Analyser. Two slices of 2 cm thickness from the centre of each loaf were analysed by the texture analyser using a probe of 1.5 inch diameter, a compression depth of 5 mm (25%) and a rate of compression of 0.5 mm/sec. In the table the average is shown of two measurements.

Crust colour was judged by a qualified baker according to the scale depicted in Table 5. As a reference the standard recipe for Dutch tin bread was used.

Crumb colour was judged by a qualified baker according to the scale depicted in Table 5. The colour of the crumb of the control breads was judged as normal (3). As a positive control the breads of 2 objects are used with the same composition as the control plus 0.5% soya flour. The proofing and baking procedure are the same as that of the control without soya flour. The latter is judged as "excellent".

The overhanging top of the bread was judged by the hanging of the top in relation to the baking tin, the lower the edges of the top the lower the judgement. The less hanging, the better the judgement.

Staling of the bread was judged by feeling the firmness of the crumb of slices of the bread. Before slicing took place, the bread was stored in a plastic bag at room temperature for 4 days. The softer the crumb of the slices is, the better the judgement.

20

Table 6. Baking performance of the lipolytic enzymes of the invention

Lipolytic enzyme	Parameter								
	Volume (%)	Dough stickyness	Dough extensibility	Dough stability	Crumb structure	Crust colour	Crumb colour	Overhanging top	Staling
NBE028	100	3	3	4	2	3	3	3	4
NBE029	104	3	3	4	3	4	4	4	3
NBE030	107	3	2	4	4	4	4	3	3
NBE031	102	3	2	4	5	4	4	4	4
NBE032	98	3	3	4	2	3	3	3	3

NBE033	105	3	2	4	2	4	3	3	3
NBE034	104	3	3	4	4	4	4	4	3
NBE036	100	3	3	4	3	4	4	4	3
NBE038	109	3	3	4	5	4	4	3	3
NBE039	109	3	3	4	4	3	4	3	3
NBE043	106	3	3	4	3	4	4	3	3
NBE045	110	3	3	4	4	3	4	4	4
NBE042	110	3	4	4	4	3	4	3	3

EXAMPLE 5

Baking experiments 2 -- batard

5 The baking performance of lipolytic enzymes according to the invention was tested in the French type of bread called "batard". Preparation of batards in a standard baking process was done by mixing 3000 g of wheat flour at circa 20°C, 70 g compressed yeast, 60 g salt, 68 ppm ascorbic acid, 30 ppm Bakezyme® HS₂₀₀₀ (fungal hemicellulase), 7 ppm Bakezyme® P500 (fungal α-amylase) and 1680 ml water (8-10°C) in a spiral mixer (Diosna: 2 minutes in speed 1; 100 Wh input in speed 2). The dough temperature was 27°C. The machineability of the dough was analysed by hand by a baker. The dough was given a bulk proof of 15 minutes in a proofing cabinet at 32°C and 90% RH. Afterwards the dough was divided into 6 pieces of 350 g, rounded and proofed for 15 minutes at 32°C and 90% RH. At the end of this period the dough pieces were moulded and shaped and given a final proof of 90 minutes at 32°C and 90% RH. The fully proofed doughs were cut in the length of the dough piece and baked in an oven at 240°C for 30 minutes with initial steam addition. After cooling down to room temperature the volumes of the loaves were determined by the BVM-method (see example 4).

20 Break, shred and shape of the breads were analysed directly after cooling down to room temperature by a qualified baker using the score in Table 7. After 16 hours (overnight) storage in a closed box at room temperature the crumb quality was assessed a qualified baker. The value for the breads (Table 8) was derived from 1 object.

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Table 7

Effect		Score				
		1	2	3	4	5
Break and shred		extremely weak and soft	weak and soft	control bread	thin and crispy crust firm break of the cut	crust too thin, too hard
Crumb structure		poor	not uniform	control bread	good	excellent
shape	height	flat	medium	control bread	larger than (3)	Much larger than (3)
	cut	cut closed	cut closed	control bread	completely opened	completely opened; teared

5

Table 8. Baking performance of the lipolytic enzymes of the invention

lipolytic enzyme	parameter				
	Dosage*	Loaf volume (%)	Break & Shred	Shape	Crumb structure
None	0	100	3	3	3
NBE028	0.75	3	4	4	4
NBE030	3	103	4	4	3
NBE031	2.5	95	4	4	3
NBE036	ND	88	3	3	3
NBE038	30	100	4	4	3

- 10. A vector according to claim 9 wherein said polynucleotide sequence according to claims 1 to 8 is operatively linked with regulatory sequences suitable for expression of said polynucleotide sequence in a suitable host cell.
- 5 11. A vector according to claim 10 wherein said suitable host cell is a filamentous fungus.
- 12. A method for manufacturing a polynucleotide according to claims 1 – 8 or a vector according to claims 9 to 11 comprising the steps of culturing a host cell transformed with said polynucleotide or said vector and isolating said polynucleotide or said vector from said host cell.
- 10 13. An isolated lipolytic enzyme selected from the group consisting of SEQ ID NO: 3, 6, 9, 12, 15, 18, 21, 24, 27, 30, 33, 36 and 39 or functional equivalents thereof.
- 14. An isolated lipolytic enzyme according to claim 13 obtainable from *Aspergillus niger*.
- 15 15. An isolated lipolytic enzyme obtainable by expressing a polynucleotide according to claims 1 to 8 or a vector according to claims 9 to 11 in an appropriate host cell, e.g. *Aspergillus niger*.
- 16. Recombinant lipolytic enzyme comprising a functional domain of any of the lipolytic enzymes according to claims 13-15.

- 20 17. A method for manufacturing a lipolytic enzyme according to claims 13 to 16 comprising the steps of transforming a suitable host cell with an isolated polynucleotide according to claims 1 to 8 or a vector according to claims 9 to 11, culturing said cell under conditions allowing expression of said polynucleotide and optionally purifying the encoded polypeptide from said cell or culture medium.
- 25 18. A recombinant host cell comprising a polynucleotide according to claims 1 to 8 or a vector according to claims 9 to 11.

- 19. A recombinant host cell expressing a lipolytic enzyme according to claims 13 to 16.
- 20. Purified antibodies reactive with a lipolytic enzyme according to claims 13 to 16.
- 21. Fusion protein comprising a lipolytic enzyme sequence according to claims 13 to 16.
- 5 22. A process for the production of dough comprising adding a lipolytic enzyme according to anyone of claims 13-16.
- 23. A process for the production of a baked product from a dough as prepared by the process of claim 22.
- 10 24. Use of a lipolytic enzyme according to anyone of claims 13-16 for the preparation of a dough and/or the baked product thereof.

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gca att gcg gta aaa ata gaa cag cca gga ata aat cca aat ccc aca 96
Ala Ile Ala Val Lys Ile Glu Gln Pro Gly Ile Asn Pro Asn Pro Thr
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gct act gta cga aat ggc acc tac tat ggt ctc cat aac cag cac tat 144
Ala Thr Val Arg Asn Gly Thr Tyr Tyr Gly Leu His Asn Gln His Tyr
35 40 45
aat caa gac ctc ttt ctc ggt att cca tat gca cag caa cct att ggt 192
Asn Gln Asp Leu Phe Leu Gly Ile Pro Tyr Ala Gln Gln Pro Ile Gly
50 55 60
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Asp Leu Arg Leu Arg Thr Pro Arg Ser Met Asn Thr Ser Trp Pro Val
65 70 75 80
cca aga aat gca aca gaa tat tca ccc gca tgt gtt gga ttt aat cag 288
Pro Arg Asn Ala Thr Glu Tyr Ser Pro Ala Cys Val Gly Phe Asn Gln
85 90 95
aca gag ggt gct tcc gaa gcc tgc ctt act ctc aat gtc gtc cgc ccg 336
Thr Glu Gly Ala Ser Glu Ala Cys Leu Thr Leu Asn Val Val Arg Pro
100 105 110
gca agc atc gct ctt tct gaa agt ctt ccc gtt gct gtc tgg att cat 384
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 Ser Phe Ile Val Asp Gln Ser Val Gln Met Glu Lys Pro Val Ile Ala
 145 150 155 160

gtc agt cta aat tat cgt ctt caa tgc tgg ggt ttt atg tgg agc aag 528
 Val Ser Leu Asn Tyr Arg Leu Gln Cys Trp Gly Phe Met Trp Ser Lys
 165 170 175

gag atg aag gaa gcc gga gta ggg aac ctg gga ctt aga gac caa cga 576
 Glu Met Lys Glu Ala Gly Val Gly Asn Leu Gly Leu Arg Asp Gln Arg
 180 185 190

tta gct ctg cat tgg ata caa gaa aac att gct gcg ttt ggt gga gac 624
 Leu Ala Leu His Trp Ile Gln Glu Asn Ile Ala Ala Phe Gly Gly Asp
 195 200 205

cct gct cag gtt aca att tgg ggt gaa agt gcc ggc gct aat agt gtt 672
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 210 215 220

ggc aca cat ctg gtt gct tac gga ggg cgc gat gat ggt ata ttc cgt 720
 Gly Thr His Leu Val Ala Tyr Gly Gly Arg Asp Asp Gly Ile Phe Arg
 225 230 235 240

gca gct atc agt gaa agt ggt gcc cca agt gtt tac caa cgt tat cca 768
 Ala Ala Ile Ser Glu Ser Gly Ala Pro Ser Val Tyr Gln Arg Tyr Pro
 245 250 255

aca cct gct gaa tgg cag ccc tat tat gat ggt att gtg aat gca tca 816
 Thr Pro Ala Glu Trp Gln Pro Tyr Tyr Asp Gly Ile Val Asn Ala Ser
 260 265 270

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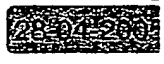
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 305 310 315 320

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370	375	380	
cta tac cca gac att cct cag ata gga atc ccc gcc ata atg gtt gga			1200
Leu Tyr Pro Asp Ile Pro Gln Ile Gly Ile Pro Ala Ile Met Val Gly			
385	390	395	400
agg cca ccg tcc gga tat gga aat caa tac aag cgt gtg gcc gca ttt			1248
Arg Pro Pro Ser Gly Tyr Gly Asn Gln Tyr Lys Arg Val Ala Ala Phe			
	405	410	415
cag ggt gat gtt aac atc cat gcc gca cgt agg ttg acc agt cag atc			1296
Gln Gly Asp Val Asn Ile His Ala Ala Arg Arg Leu Thr Ser Gln Ile			
	420	425	430
tgg tca tcc cgc aat atc tca gta tat agc tac atg ttt gac gtt atc			1344
Trp Ser Ser Arg Asn Ile Ser Val Tyr Ser Tyr Met Phe Asp Val Ile			
	435	440	445
agc cct gga tat ggc ccc tct gct ggt tcc tat gct ggg gct act cat			1392
Ser Pro Gly Tyr Gly Pro Ser Ala Gly Ser Tyr Ala Gly Ala Thr His			
	450	455	460
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Gly Thr Asp Ile Pro Tyr Val Phe Tyr Asn Leu Asp Gly Leu Gly Tyr			
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gac tcg aac aac aag tcc ata gaa agc ata cct aac agt tat tcc cgc			1488
Asp Ser Asn Asn Lys Ser Ile Glu Ser Ile Pro Asn Ser Tyr Ser Arg			
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Met Ser Lys Ile Met Ser Arg Met Trp Val Ser Phe Val Thr Thr Leu			
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gac cca aat cat tct gga ggt atg gtc cca cat ccc att cct atg att			1584
Asp Pro Asn His Ser Gly Gly Met Val Pro His Pro Ile Pro Met Ile			
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Val Gln Trp Pro Pro Tyr Asn Ile Asp Asn Pro Glu Ile Ile Phe Phe			
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Asp Thr Asp Val Thr Asn Leu Thr Tyr Thr Trp Pro Ala Gly Leu Tyr			
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Pro Arg Asn Ala Thr Glu Tyr Ser Pro Ala Cys Val Gly Phe Asn Gln
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100 105 110
Ala Ser Ile Ala Leu Ser Glu Ser Leu Pro Val Ala Val Trp Ile His
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130 135 140
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145 150 155 160
Val Ser Leu Asn Tyr Arg Leu Gln Cys Trp Gly Phe Met Trp Ser Lys
165 170 175
Glu Met Lys Glu Ala Gly Val Gly Asn Leu Gly Leu Arg Asp Gln Arg
180 185 190
Leu Ala Leu His Trp Ile Gln Glu Asn Ile Ala Ala Phe Gly Gly Asp
195 200 205

Pro Ala Gln Val Thr Ile Trp Gly Glu Ser Ala Gly Ala Asn Ser Val
210 215 220
Gly Thr His Leu Val Ala Tyr Gly Gly Arg Asp Asp Gly Ile Phe Arg
225 230 235 240
Ala Ala Ile Ser Glu Ser Gly Ala Pro Ser Val Tyr Gln Arg Tyr Pro
245 250 255
Thr Pro Ala Glu Trp Gln Pro Tyr Tyr Asp Gly Ile Val Asn Ala Ser
260 265 270
Gly Cys Ser Ser Ala Thr Asp Thr Leu Ala Cys Leu Arg Thr Ile Pro
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Leu Gly Thr Tyr Ala Pro Tyr Tyr Ala Asn Leu Thr Trp Glu Gln Pro	
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Arg Thr Leu Ser Asn Trp Ser Asn Leu Thr Val Glu Thr Arg Thr Gly	
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Thr Phe Ile Gly Met Leu Asn Asp Thr Tyr Pro Asp Val Arg Gln Phe	
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Leu Arg Val Pro Tyr Ala Lys Pro Pro Ile Gly Asp Leu Arg Trp Leu	
65 70 75 80	
cct cct cat cgg ctt gac aac tca agc aga aca tat gac tcc acc ttc	288
Pro Pro His Arg Leu Asp Asn Ser Ser Arg Thr Tyr Asp Ser Thr Phe	
85 90 95	
tat ggc cca gcc tgt ccg cag tat gtt cca gca gag agc gat ttt tgg	336
Tyr Gly Pro Ala Cys Pro Gln Tyr Val Pro Ala Glu Ser Asp Phe Trp	
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Asn Glu Tyr Glu Pro Glu Asn Leu Leu Leu Asn Val Gly Glu Arg Leu	
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Asn Gln Gly Ser Thr Ala Trp Ser Ser Ser Glu Asp Cys Leu Ser Leu	
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165	170 175
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Pro Ser Gln Leu Pro Ser Ala Trp Val Ser Arg Ser Gln Glu His Ile	
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Val Val Thr Ile Asn Tyr Arg Val Asn Ile Phe Gly Asn Pro Lys Ser	
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Arg Ala Leu Asn Asp Thr Ser Leu Thr Leu Met Asp Val Arg Ala Ala	
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Val Glu Trp Val Tyr Glu Asn Ile Glu Ala Phe Gly Gly Asn Pro Glu	
225	230 235 240
aat att atg gtc aga cta caa gtt tcc tct cac atg act aga gct aac	768
Asn Ile Met Val Arg Leu Gln Val Ser Ser His Met Thr Arg Ala Asn	
245	250 255
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Ser Lys Gln Leu Trp Gly Gln Ser Gln Gly Ala Leu Leu Thr His Leu	
260	265 270
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Tyr Thr Leu Ala Trp Pro Glu Glu Pro Leu Ala Ala Lys Phe Gly Val	
275	280 285
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Ile Ser Gln Gly Ala Ser Ala Thr Leu Asn Leu Ser Thr Thr Pro Asp	
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Val Tyr Gln Asp Phe Asp Ile Val Ala Lys Gly Leu Gly Cys Asn Tyr	
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Gly Asp Asp Ala Glu Ala Glu Leu Glu Cys Met Arg Gly Ile Ser Trp	
325	330 335
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Val Gln Ile Glu Glu Tyr Ile Asn Arg Tyr Asn Ser Ser Pro Ser Ile	
340	345 350
gct ttc acg aac tat att ccc gat gag aaa tac atc ttc tcc gac gaa	1104
Ala Phe Thr Asn Tyr Ile Pro Asp Glu Lys Tyr Ile Phe Ser Asp Glu	
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Arg Gln Arg Tyr Leu Glu Arg Lys Val Ala Arg Gly Pro Ser Ile Arg	
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- <212> DNA
- <213> *Aspergillus niger*

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agtggccctg	taccagaca	ccaccagta	cgggtctccg	ttcaggacag	gcgcggccaa	2160
caactggtat	ccgaattta	agcgattggc	cgccattctc	ggcgacttgg	tcttaccat	2220
taccggcgg	gcattcctct	cgtatgcaga	ggaaatctcc	cctgatcttc	cgaactggtc	2280
gtacctggcg	acctatgact	atggcacccc	agttctgggg	accttccacg	gaagtgacct	2340
gctgcaggtg	ttctatggga	tcaagccaaa	ctatgcagct	agttctagcc	acacgtacta	2400
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gcagtggaag	gaatcgcggc	agttgatgaa	tttcggagcg	aacgacgcca	gtctccttac	2520
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aagagcttga	agtggcgaga	tgtctctgca	ggaattcaag	ctagatgcta	agcgatattg	3060
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- <210> 11
- <211> 1689
- <212> DNA
- <213> *Aspergillus niger*
- <220>



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<221> CDS

<222> (1)..(1689)

<400> 11

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tgg gcg act cca gtt caa cgg gat gca gct cct act gtc act att gcg	96
Trp Ala Thr Pro Val Gln Arg Asp Ala Ala Pro Thr Val Thr Ile Ala	
20 25 30	
cat cca tcg gcc acc gtc att gga aaa tct ggc aat gtc gag agc ttc	144
His Pro Ser Ala Thr Val Ile Gly Lys Ser Gly Asn Val Glu Ser Phe	
35 40 45	
aac aat att ccc ttt gcg cag gcc ccc aca ggc tcg ctg cgt ctg aag	192
Asn Asn Ile Pro Phe Ala Gln Ala Pro Thr Gly Ser Leu Arg Leu Lys	
50 55 60	
ccc cca caa ccc ttg gaa act gcc ctc ggc act gtt cag gcc aca gga	240
Pro Pro Gln Pro Leu Glu Thr Ala Leu Gly Thr Val Gln Ala Thr Gly	
65 70 75 80	
gcc tcg caa tcg tgt ccg cag atg tac ttc acc acg gat gag agc gaa	288
Ala Ser Gln Ser Cys Pro Gln Met Tyr Phe Thr Thr Asp Glu Ser Glu	
85 90 95	
ttc ccg aca tcg gtc att ggc ctc ctc gct gat ctc cct ttg gta cag	336
Phe Pro Thr Ser Val Ile Gly Leu Leu Ala Asp Leu Pro Leu Val Gln	
100 105 110	
tcg gct acc aat gct ctc gag gat tgc ctg aac att gac att cgg cgt	384
Ser Ala Thr Asn Ala Leu Glu Asp Cys Leu Asn Ile Asp Ile Arg Arg	
115 120 125	
ccg gcc ggg acc acc gcg gac tcg aag ctg cct gtg ctg gtc tgg atc	432
Pro Ala Gly Thr Thr Ala Asp Ser Lys Leu Pro Val Leu Val Trp Ile	
130 135 140	
ttt ggc gga ggc ttt gaa ctt ggt tca aag gcg atg tat gat ggt aca	480
Phe Gly Gly Gly Phe Glu Leu Gly Ser Lys Ala Met Tyr Asp Gly Thr	
145 150 155 160	
acg atg gta tca tcg tcg ata gac aag aac atg cct atc gtg ttt gta	528
Thr Met Val Ser Ser Ser Ile Asp Lys Asn Met Pro Ile Val Phe Val	
165 170 175	
gca atg aat tat cgc gtg gga ggt ttc ggg ttc ttg ccc gga aag gag	576
Ala Met Asn Tyr Arg Val Gly Gly Phe Gly Phe Leu Pro Gly Lys Glu	
180 185 190	
atc ctg gag gac ggg tcc gcg aac cta ggg ctc ctg gac caa cgc ctt	624
Ile Leu Glu Asp Gly Ser Ala Asn Leu Gly Leu Leu Asp Gln Arg Leu	
195 200 205	
gcc ctg cag tgg gtt gcc gac aac atc gag gcc ttt ggt gga gac ccg	672

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Ala Leu Gln Trp Val Ala Asp Asn Ile Glu Ala Phe Gly Gly Asp Pro
 210 215 220
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 Asp Lys Val Thr Ile Trp Gly Glu Ser Ala Gly Ala Ile Ser Val Phe
 225 230 235 240
 gat cag atg atc ttg tac gac gga aac atc act tac aag gat aag ccc 768
 Asp Gln Met Ile Leu Tyr Asp Gly Asn Ile Thr Tyr Lys Asp Lys Pro
 245 250 255
 ttg ttc cgg ggg gcc atc atg gac tcc ggt agt gtt gtt ccc gca gac 816
 Leu Phe Arg Gly Ala Ile Met Asp Ser Gly Ser Val Val Pro Ala Asp
 260 265 270
 ccc gtc gat ggg gtc aag gga cag caa gta tat gat gcg gta gtg gaa 864
 Pro Val Asp Gly Val Lys Gly Gln Gln Val Tyr Asp Ala Val Val Glu
 275 280 285
 tct gca ggc tgt tcc tct tct aac gac acc cta gct tgt ctg cgt gaa 912
 Ser Ala Gly Cys Ser Ser Ser Asn Asp Thr Leu Ala Cys Leu Arg Glu
 290 295 300
 cta gac tac acc gac ttc ctc aat gcg gca aac tcc gtg cca ggc att 960
 Leu Asp Tyr Thr Asp Phe Leu Asn Ala Ala Asn Ser Val Pro Gly Ile
 305 310 315 320
 tta agc tac cat tct gtg gcg tta tca tat gtg cct cga ccg gac ggg 1008
 Leu Ser Tyr His Ser Val Ala Leu Ser Tyr Val Pro Arg Pro Asp Gly
 325 330 335
 acg gcg ttg tcg gca tca ccg gac gtt ttg ggc aaa gca ggg aaa tat 1056
 Thr Ala Leu Ser Ala Ser Pro Asp Val Leu Gly Lys Ala Gly Lys Tyr
 340 345 350
 gct cgg gtc ccg ttc atc gtg ggc gac caa gag gat gag ggg acc tta 1104
 Ala Arg Val Pro Phe Ile Val Gly Asp Gln Glu Asp Glu Gly Thr Leu
 355 360 365
 ttc gcc ttg ttt cag tcc aac att acg acg atc gac gag gtg gtc gac 1152
 Phe Ala Leu Phe Gln Ser Asn Ile Thr Thr Ile Asp Glu Val Val Asp
 370 375 380
 tac ctg gcc tca tac ttc ttc tat gac gct agc cga gag cag ctt gaa 1200
 Tyr Leu Ala Ser Tyr Phe Phe Tyr Asp Ala Ser Arg Glu Gln Leu Glu
 385 390 395 400
 gaa cta gtg gcc ctg tac cca gac acc acc acg tac ggg tct ccg ttc 1248
 Glu Leu Val Ala Leu Tyr Pro Asp Thr Thr Thr Tyr Gly Ser Pro Phe
 405 410 415
 agg aca ggc gcg gcc aac aac tgg tat ccg caa ttt aag cga ttg gcc 1296
 Arg Thr Gly Ala Ala Asn Asn Trp Tyr Pro Gln Phe Lys Arg Leu Ala
 420 425 430
 gcc att ctc ggc gac ttg gtc ttc acc att acc cgg cgg gca ttc ctc 1344
 Ala Ile Leu Gly Asp Leu Val Phe Thr Ile Thr Arg Arg Ala Phe Leu
 435 440 445

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tcg tat gca gag gaa atc tcc cct gat ctt ccg aac tgg tcg tac ctg 1392
 Ser Tyr Ala Glu Glu Ile Ser Pro Asp Leu Pro Asn Trp Ser Tyr Leu
 450 455 460

gcg acc tat gac tat ggc acc cca gtt ctg ggg acc ttc cac gga agt 1440
 Ala Thr Tyr Asp Tyr Gly Thr Pro Val Leu Gly Thr Phe His Gly Ser
 465 470 475 480

gac ctg ctg cag gtg ttc tat ggg atc aag cca aac tat gca gct agt 1488
 Asp Leu Leu Gln Val Phe Tyr Gly Ile Lys Pro Asn Tyr Ala Ala Ser
 485 490 495

tct agc cac acg tac tat ctg agc ttt gtg tat acg ctg gat ccg aac 1536
 Ser Ser His Thr Tyr Tyr Leu Ser Phe Val Tyr Thr Leu Asp Pro Asn
 500 505 510

tcc aac cgg ggg gag tac att gag tgg ccg cag tgg aag gaa tcg cgg 1584
 Ser Asn Arg Gly Glu Tyr Ile Glu Trp Pro Gln Trp Lys Glu Ser Arg
 515 520 525

cag ttg atg aat ttc gga gcg aac gac gcc agt ctc ctt acg gat gat 1632
 Gln Leu Met Asn Phe Gly Ala Asn Asp Ala Ser Leu Leu Thr Asp Asp
 530 535 540

ttc cgc aac ggg aca tat gag ttc atc ctg cag aat acc gcg gcg ttc 1680
 Phe Arg Asn Gly Thr Tyr Glu Phe Ile Leu Gln Asn Thr Ala Ala Phe
 545 550 555 560

cac atc tga 1689
 His Ile

- <210> 12
- <211> 562
- <212> PRT
- <213> Aspergillus niger

<400> 12

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 20 25 30

His Pro Ser Ala Thr Val Ile Gly Lys Ser Gly Asn Val Gln Ser Phe
 35 40 45

Asn Asn Ile Pro Phe Ala Gln Ala Pro Thr Gly Ser Leu Arg Leu Lys
 50 55 60

Pro Pro Gln Pro Leu Glu Thr Ala Leu Gly Thr Val Gln Ala Thr Gly
 65 70 75 80

Ala Ser Gln Ser Cys Pro Gln Met Tyr Phe Thr Thr Asp Gln Ser Glu
 85 90 95

Phe Pro Thr Ser Val Ile Gly Leu Leu Ala Asp Leu Pro Leu Val Gln

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21078WO.ST25.txt

100 105 110
 Ser Ala Thr Asn Ala Leu Glu Asp Cys Leu Asn Ile Asp Ile Arg Arg
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 Pro Ala Gly Thr Thr Ala Asp Ser Lys Leu Pro Val Leu Val Trp Ile
 130 135 140
 Phe Gly Gly Gly Phe Glu Leu Gly Ser Lys Ala Met Tyr Asp Gly Thr
 145 150 155 160
 Thr Met Val Ser Ser Ser Ile Asp Lys Asn Met Pro Ile Val Phe Val
 165 170 175
 Ala Met Asn Tyr Arg Val Gly Gly Phe Gly Phe Leu Pro Gly Lys Glu
 180 185 190
 Ile Leu Glu Asp Gly Ser Ala Asn Leu Gly Leu Leu Asp Gln Arg Leu
 195 200 205
 Ala Leu Gln Trp Val Ala Asp Asn Ile Glu Ala Phe Gly Gly Asp Pro
 210 215 220
 Asp Lys Val Thr Ile Trp Gly Glu Ser Ala Gly Ala Ile Ser Val Phe
 225 230 235 240
 Asp Gln Met Ile Leu Tyr Asp Gly Asn Ile Thr Tyr Lys Asp Lys Pro
 245 250 255
 Leu Phe Arg Gly Ala Ile Met Asp Ser Gly Ser Val Val Pro Ala Asp
 260 265 270
 Pro Val Asp Gly Val Lys Gly Gln Gln Val Tyr Asp Ala Val Val Glu
 275 280 285
 Ser Ala Gly Cys Ser Ser Ser Asn Asp Thr Leu Ala Cys Leu Arg Glu
 290 295 300
 Leu Asp Tyr Thr Asp Phe Leu Asn Ala Ala Asn Ser Val Pro Gly Ile
 305 310 315 320
 Leu Ser Tyr His Ser Val Ala Leu Ser Tyr Val Pro Arg Pro Asp Gly
 325 330 335
 Thr Ala Leu Ser Ala Ser Pro Asp Val Leu Gly Lys Ala Gly Lys Tyr
 340 345 350
 Ala Arg Val Pro Phe Ile Val Gly Asp Gln Glu Asp Glu Gly Thr Leu
 355 360 365

Phe Ala Leu Phe Gln Ser Asn Ile Thr Thr Ile Asp Glu Val Val Asp
 370 375 380
 Tyr Leu Ala Ser Tyr Phe Phe Tyr Asp Ala Ser Arg Glu Gln Leu Glu
 385 390 395 400
 Glu Leu Val Ala Leu Tyr Pro Asp Thr Thr Thr Tyr Gly Ser Pro Phe
 405 410 415
 Arg Thr Gly Ala Ala Asn Asn Trp Tyr Pro Gln Phe Lys Arg Leu Ala
 420 425 430
 Ala Ile Leu Gly Asp Leu Val Phe Thr Ile Thr Arg Arg Ala Phe Leu
 435 440 445
 Ser Tyr Ala Glu Glu Ile Ser Pro Asp Leu Pro Asn Trp Ser Tyr Leu

21078\0.ST25.txt

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cgaggtgaaa gaaaagacgg gcgccgacaa gattgacctt gtcggctact ccgaaggcgc 1500
cttccagacc ctctacgtcc ctaagttcga ggatggatc tcggagatgc tggataagct 1560
ggtggccatt gcacctccca ccagaggcac caactggcg gggatctatg acatgcata 1620
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cggcaggtga gactatcacc ttctgaaaat ttgtatataa gcatttatat ttggataacc 2040
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<210> 14
<211> 834
<212> DNA
<213> Aspergillus niger

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<220>
<221> CDS
<222> (1)..(834)

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tta ggc tac agc atc aac gac ttc tcc tgc aat agc acc gaa cac ccg 96
Leu Gly Tyr Ser Ile Asn Asp Phe Ser Cys Asn Ser Thr Glu His Pro
20 25 30
aat cca gtt gtg ctc cta cat ggg cta ggc gcc acc tac tac gaa gac 144
Asn Pro Val Val Leu Leu His Gly Leu Gly Ala Thr Tyr Tyr Glu Asp
35 40 45
ttg aat tac ctg caa ggt tgg cta cag acc caa ggc tat tgc act tac 192
Leu Asn Tyr Leu Gln Gly Trp Leu Gln Thr Gln Gly Tyr Cys Thr Tyr
50 55 60
gcc aaa acc tac ggt gca tat gaa ggc ttc ccc ttt gtc ggc gcc ctc 240
Ala Lys Thr Tyr Gly Ala Tyr Glu Gly Phe Pro Phe Val Gly Gly Leu
65 70 75 80
aag gcc atc gcc gaa tcg gcc acg gaa atc gcc gcg tac atc cgc gag 288
Lys Ala Ile Ala Glu Ser Ala Thr Glu Ile Ala Ala Tyr Ile Arg Glu
85 90 95
gtg aaa gaa aag acg ggc gcc gac aag att gac ctt gtc ggt cac tcc 336

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Val	Lys	Glu	Lys	Thr	Gly	Ala	Asp	Lys	Ile	Asp	Leu	Val	Gly	His	Ser	
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Glu	Gly	Ala	Phe	Gln	Thr	Leu	Tyr	Val	Pro	Lys	Phe	Glu	Asp	Gly	Ile	
			115				120					125				
tcg	gag	atg	ctg	gat	aag	ctg	gtg	gcc	att	gca	cct	ccc	acc	aga	ggc	432
Ser	Glu	Met	Leu	Asp	Lys	Leu	Val	Ala	Ile	Ala	Pro	Pro	Thr	Arg	Gly	
			130			135				140						
acc	aac	ttg	gcg	ggg	atc	tat	gac	atc	gca	tat	ggt	ctg	gga	aat	cta	480
Thr	Asn	Leu	Ala	Gly	Ile	Tyr	Asp	Ile	Ala	Tyr	Val	Leu	Gly	Asn	Leu	
			145		150				155					160		
tcg	cgc	gat	ctg	ata	ggc	gac	gtc	ctg	gat	acc	gtg	ggc	tgc	gcc	gcc	528
Ser	Arg	Asp	Leu	Ile	Gly	Asp	Val	Leu	Asp	Thr	Val	Gly	Cys	Ala	Ala	
				165				170					175			
tgt	gat	gat	ctg	ggt	ccg	gat	gga	gca	gcg	att	gac	cgc	ttg	aac	gat	576
Cys	Asp	Asp	Leu	Gly	Pro	Asp	Gly	Ala	Ala	Ile	Asp	Arg	Leu	Asn	Asp	
			180				185						190			
ggc	gag	cct	atc	gtg	cag	ccg	gga	aat	aat	cta	acg	gtg	att	gca	tcg	624
Gly	Glu	Pro	Ile	Val	Gln	Pro	Gly	Asn	Asn	Leu	Thr	Val	Ile	Ala	Ser	
			195				200					205				
cgg	tcc	gac	gaa	ttg	gtc	acc	cca	acc	acc	acc	tcc	ttc	gtg	cat	gaa	672
Arg	Ser	Asp	Glu	Leu	Val	Thr	Pro	Thr	Thr	Thr	Ser	Phe	Val	His	Glu	
			210			215					220					
gat	ggg	gtg	acc	aat	gaa	tgg	gtg	caa	gac	act	tgt	cct	cta	gac	cct	720
Asp	Gly	Val	Thr	Asn	Glu	Trp	Val	Gln	Asp	Thr	Cys	Pro	Leu	Asp	Pro	
				225		230				235				240		
gtc	ggt	cat	atc	ggt	gag	gca	tac	gat	ctg	aac	gtc	tgg	aat	ttg	gtc	768
Val	Gly	His	Ile	Gly	Glu	Ala	Tyr	Asp	Leu	Asn	Val	Trp	Asn	Leu	Val	
				245				250					255			
aaa	aac	gcc	ttg	gac	tct	acg	ccg	aag	cgt	gag	ttc	gtc	tgc	tcg	ctg	816
Lys	Asn	Ala	Leu	Asp	Ser	Thr	Pro	Lys	Arg	Glu	Phe	Val	Cys	Ser	Leu	
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gga	tct	ccc	ggc	agg	tga											834
Gly	Ser	Pro	Gly	Arg												
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- <210> 15
- <211> 277
- <212> PRT
- <213> Aspergillus niger

<400> 15
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21078WO.ST25.txt

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Leu Asn Tyr Leu Gln Gly Trp Leu Gln Thr Gln Gly Tyr Cys Thr Tyr
                50           55           60
Ala Lys Thr Tyr Gly Ala Tyr Glu Gly Phe Pro Phe Val Gly Gly Leu
                65           70           75           80
Lys Ala Ile Ala Glu Ser Ala Thr Glu Ile Ala Ala Tyr Ile Arg Glu
                85           90           95
Val Lys Glu Lys Thr Gly Ala Asp Lys Ile Asp Leu Val Gly His Ser
                100          105          110
Glu Gly Ala Phe Gln Thr Leu Tyr Val Pro Lys Phe Glu Asp Gly Ile
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Ser Glu Met Leu Asp Lys Leu Val Ala Ile Ala Pro Pro Thr Arg Gly
                130          135          140
Thr Asn Leu Ala Gly Ile Tyr Asp Ile Ala Tyr Val Leu Gly Asn Leu
                145          150          155          160
Ser Arg Asp Leu Ile Gly Asp Val Leu Asp Thr Val Gly Cys Ala Ala
                165          170          175
Cys Asp Asp Leu Gly Pro Asp Gly Ala Ala Ile Asp Arg Leu Asn Asp
                180          185          190
Gly Glu Pro Ile Val Gln Pro Gly Asn Asn Leu Thr Val Ile Ala Ser
                195          200          205
Arg Ser Asp Glu Leu Val Thr Pro Thr Thr Thr Ser Phe Val His Glu
                210          215          220
Asp Gly Val Thr Asn Glu Trp Val Gln Asp Thr Cys Pro Leu Asp Pro
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Val Gly His Ile Gly Glu Ala Tyr Asp Leu Asn Val Trp Asn Leu Val
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Gly Ser Pro Gly Arg
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- <210> 16
- <211> 1881
- <212> DNA
- <213> Aspergillus niger

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<400> 16 .
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21078WO.ST25.txt

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aatgactact ggcacatctg c 1881

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<210> 17
 <211> 1257
 <212> DNA
 <213> *Aspergillus niger*

<220>
 <221> CDS
 <222> (1)..(1257)

<400> 17
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21078WO.ST25.txt

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Ala Thr Ala Leu Pro Thr Pro Gly Ser Thr Pro Ile Pro Pro Ser Gln
          20          25          30
gat ccc tgg tac agt gcg ccc gag ggc ttc gag gag gct gat ccc ggt      144
Asp Pro Trp Tyr Ser Ala Pro Glu Gly Phe Glu Glu Ala Asp Pro Gly
          35          40          45
gcc atc ctg cgc gtg cgg ccc gcg ccc ggc aac ttg acc gtg gta gtg      192
Ala Ile Leu Arg Val Arg Pro Ala Pro Gly Asn Leu Thr Val Val Val
          50          55          60
ggc aat gcg tcg gcg gcc tac aac atc ctc tac cgc act aca gac agt      240
Gly Asn Ala Ser Ala Ala Tyr Asn Ile Leu Tyr Arg Thr Thr Asp Ser
          65          70          75          80
cag tac aag ccc tcc tgg gct gtg acc acc ctg ctg gtg ccc ccc gtg      288
Gln Tyr Lys Pro Ser Trp Ala Val Thr Thr Leu Leu Val Pro Pro Val
          85          90          95
gcc gcc tcc gcc gcc gtc aac cag agt gtc ctg ctc tcc cac cag atc      336
Ala Ala Ser Ala Ala Val Asn Gln Ser Val Leu Leu Ser His Gln Ile
          100          105          110
gcc tac gat tcg ttc gac gtc aat gcc agt ccc agc tac gcc atg tac      384
Ala Tyr Asp Ser Phe Asp Val Asn Ala Ser Pro Ser Tyr Ala Met Tyr
          115          120          125
acc agc ccg ccc tcc gat att atc ctc gcc ctg cag cgc ggc tgg ttc      432
Thr Ser Pro Pro Ser Asp Ile Ile Leu Ala Leu Gln Arg Gly Trp Phe
          130          135          140
gtt aac gtc ccc gat tac gag ggc ccc aat gcc tct ttc acc gcc ggt      480
Val Asn Val Pro Asp Tyr Glu Gly Pro Asn Ala Ser Phe Thr Ala Gly
          145          150          155          160
gtg cag tcc ggc cat gcc acc ctc gac tcg gtc cgc agc gtg ctc gcc      528
Val Gln Ser Gly His Ala Thr Leu Asp Ser Val Arg Ser Val Leu Ala
          165          170          175
tcc gga ttc ggc etg aac gag gac gcc cag tac gct etg tgg ggt tac 576
Ser Gly Phe Gly Leu Asn Glu Asp Ala Gln Tyr Ala Leu Trp Gly Tyr
          180          185          190
tct ggc ggt gcc ttg gcc agc gaa tgg gct gct gaa ctg cag atg caa      624
Ser Gly Gly Ala Leu Ala Ser Glu Trp Ala Ala Glu Leu Gln Met Gln
          195          200          205
tac gct ccc gag ttg aac att gcc ggt ctg gcc gtg ggt ggt ctc act      672
Tyr Ala Pro Glu Leu Asn Ile Ala Gly Leu Ala Val Gly Gly Leu Thr
          210          215          220
ccc aat gtt acc agc gtc atg gac acg gtg acc tcg acc atc agt gcg      720
Pro Asn Val Thr Ser Val Met Asp Thr Val Thr Ser Thr Ile Ser Ala
          225          230          235          240

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gga ctc atc ccc gcc gcc gcc ctg ggt ctg tcg agc cag cac ccc gag 768
 Gly Leu Ile Pro Ala Ala Ala Leu Gly Leu Ser Ser Gln His Pro Glu
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acc tac gag ttc atc ctc agc cag ctc aag acg acg gga ccc tac aac 816
 Thr Tyr Glu Phe Ile Leu Ser Gln Leu Lys Thr Thr Gly Pro Tyr Asn
 260 265 270

cgc aca gga ttc cta gcc gcc aag gac ctg acc ctg tcc gag gcg gag 864
 Arg Thr Gly Phe Leu Ala Ala Lys Asp Leu Thr Leu Ser Glu Ala Glu
 275 280 285

gtc ttc tac gcc ttc cag aac atc ttc gat tac ttt gtc aac gga tcg 912
 Val Phe Tyr Ala Phe Gln Asn Ile Phe Asp Tyr Phe Val Asn Gly Ser
 290 295 300

gcc acg ttc cag gcg gag gtg gtg cag aag gcg ctg aac cag gac gga 960
 Ala Thr Phe Gln Ala Glu Val Val Gln Lys Ala Leu Asn Gln Asp Gly
 305 310 315 320

tac atg ggc tac cat ggg ttc ccg cag atg ccg gtg ctc gcg tac aag 1008
 Tyr Met Gly Tyr His Gly Phe Pro Gln Met Pro Val Leu Ala Tyr Lys
 325 330 335

gct att cac gat gag atc agt ccc atc cag gat acg gat cgc gtg atc 1056
 Ala Ile His Asp Glu Ile Ser Pro Ile Gln Asp Thr Asp Arg Val Ile
 340 345 350

aag cgc tac tgt ggt ctg gga ttg aac atc ttg tat gag cgg aac acc 1104
 Lys Arg Tyr Cys Gly Leu Gly Leu Asn Ile Leu Tyr Glu Arg Asn Thr
 355 360 365

atc ggt ggc cac tcg gca gag cag gtg aat ggc aac gcc agg gcg tgg 1152
 Ile Gly Gly His Ser Ala Glu Gln Val Asn Gly Asn Ala Arg Ala Trp
 370 375 380

aac tgg ttg acg agc att ttc gac gga acg tat gcg cag cag tac aag 1200
 Asn Trp Leu Thr Ser Ile Phe Asp Gly Thr Tyr Ala Gln Gln Tyr Lys
 385 390 395 400

acc gag ggg tgc acg atc cgc aat gtc act ctg aac acg act tcc tcc 1248
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 405 410 415

ggt tat tag 1257
 Val Tyr

- <210> 18
- <211> 418
- <212> PRT
- <213> Aspergillus niger

<400> 18
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Ala Ile Leu Arg Val Arg Pro Ala Pro Gly Asn Leu Thr Val Val Val
      50           55           60
Gly Asn Ala Ser Ala Ala Tyr Asn Ile Leu Tyr Arg Thr Thr Asp Ser
      65           70           75           80
Gln Tyr Lys Pro Ser Trp Ala Val Thr Thr Leu Leu Val Pro Pro Val
      85           90           95
Ala Ala Ser Ala Ala Val Asn Gln Ser Val Leu Leu Ser His Gln Ile
      100           105           110
Ala Tyr Asp Ser Phe Asp Val Asn Ala Ser Pro Ser Tyr Ala Met Tyr
      115           120           125
Thr Ser Pro Pro Ser Asp Ile Ile Leu Ala Leu Gln Arg Gly Trp Phe
      130           135           140
Val Asn Val Pro Asp Tyr Glu Gly Pro Asn Ala Ser Phe Thr Ala Gly
      145           150           155           160
Val Gln Ser Gly His Ala Thr Leu Asp Ser Val Arg Ser Val Leu Ala
      165           170           175
Ser Gly Phe Gly Leu Asn Glu Asp Ala Gln Tyr Ala Leu Trp Gly Tyr
      180           185           190
Ser Gly Gly Ala Leu Ala Ser Glu Trp Ala Ala Glu Leu Gln Met Gln
      195           200           205
Tyr Ala Pro Glu Leu Asn Ile Ala Gly Leu Ala Val Gly Gly Leu Thr
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Pro Asn Val Thr Ser Val Met Asp Thr Val Thr Ser Thr Ile Ser Ala
      225           230           235           240
Gly Leu Ile Pro Ala Ala Ala Leu Gly Leu Ser Ser Gln His Pro Glu
      245           250           255
Thr Tyr Glu Phe Ile Leu Ser Gln Leu Lys Thr Thr Gly Pro Tyr Asn
      260           265           270
Arg Thr Gly Phe Leu Ala Ala Lys Asp Leu Thr Leu Ser Glu Ala Glu
      275           280           285
Val Phe Tyr Ala Phe Gln Asn Ile Phe Asp Tyr Phe Val Asn Gly Ser
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Ala Thr Phe Gln Ala Glu Val Val Gln Lys Ala Leu Asn Gln Asp Gly
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Tyr Met Gly Tyr His Gly Phe Pro Gln Met Pro Val Leu Ala Tyr Lys
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Ala Ile His Asp Glu Ile Ser Pro Ile Gln Asp Thr Asp Arg Val Ile
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 <213> Aspergillus niger

<220>
 <221> CDS
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gcc ctt ttt ggg tcg ctg att ttg gta ttg ctg gaa tgg gtt ata cat 96
Ala Leu Phe Gly Ser Leu Ile Leu Val Leu Leu Glu Trp Val Ile His
20 25 30
att atc aca ttc tgt ctg cct gaa cct gtt att aag ttc tgt tac gat 144
Ile Ile Thr Phe Cys Leu Pro Glu Pro Val Ile Lys Phe Cys Tyr Asp
35 40 45
cga tcc aag act atc ttc aac gcc ttc att cct ccc gat gac ccg gct 192
Arg Ser Lys Thr Ile Phe Asn Ala Phe Ile Pro Pro Asp Asp Pro Ala
50 55 60
aag cgc ggt aaa gaa gag aaa att gct gcg tcg gtt gct ctg gcg tcg 240

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Asp Phe Thr Asp Ile Cys Ala Leu Phe Gly Tyr Glu Ala Glu Glu His
85 90 95
atc gtc cag aca ggg gat ggc tat ctg ctt ggt ctg cac cga ctg ccc 336
Ile Val Gln Thr Gly Asp Gly Tyr Leu Leu Gly Leu His Arg Leu Pro
100 105 110
tat cgg aaa gga gag gag ggg agg aag atc aac cag ggc gaa ggg agc 384
Tyr Arg Lys Gly Glu Glu Gly Arg Lys Ile Asn Gln Gly Glu Gly Ser
115 120 125
atc aag aag aag gtc gtc tat ctc cac cat ggt ctc atg atg tgc agt 432
Ile Lys Lys Lys Val Val Tyr Leu His His Gly Leu Met Met Cys Ser
130 135 140
gaa gtc tgg atc tgt ctg tca gag gag cag cga tgc ctt ccg ttt caa 480
Glu Val Trp Ile Cys Leu Ser Glu Glu Gln Arg Cys Leu Pro Phe Gln
145 150 155 160
tta gtc gaa agg ggc tat gac gtg tgg ttg ggg aac aat aga gga aac 528
Leu Val Glu Arg Gly Tyr Asp Val Trp Leu Gly Asn Asn Arg Gly Asn
165 170 175
aag tac tcg aag aag tcc gtc aag cat tcg ccc ctg tcg aac gag ttc 576
Lys Tyr Ser Lys Lys Ser Val Lys His Ser Pro Leu Ser Asn Glu Phe
180 185 190
tgg gac ttt tcg atc gat cag ttc tcg ttc cat gat atc cca gac agc 624
Trp Asp Phe Ser Ile Asp Gln Phe Ser Phe His Asp Ile Pro Asp Ser
195 200 205
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Ile Lys Tyr Ile Leu Glu Val Thr Gly Gln Pro Ser Leu Ser Tyr Val
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Gly Phe Ser Gln Gly Thr Ala Gln Ala Phe Ala Thr Leu Ser Ile His
225 230 235 240
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Pro Leu Leu Asn Gln Lys Ile Asp Val Phe Val Ala Leu Ala Pro Ala
245 250 255
atg gct ccg aca ggt ctt cca aat cat ctc gtg gac tcg ctc atg aag 816
Met Ala Pro Thr Gly Leu Pro Asn His Leu Val Asp Ser Leu Met Lys
260 265 270
gct tcg ccg aac ttc ctg ttt ctg ctg ttt ggc aga cgc agc atc ctt 864
Ala Ser Pro Asn Phe Leu Phe Leu Leu Phe Gly Arg Arg Ser Ile Leu
275 280 285
agc tca acg acg atg tgg cag aca att ctc tac ccg cct atc ttt gtt 912
Ser Ser Thr Thr Met Trp Gln Thr Ile Leu Tyr Pro Pro Ile Phe Val
290 295 300

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Asn Ile Ser Arg Trp Gln Lys Leu Ala Gly Tyr Leu His Leu Phe Ser	
325 330 335	
ttc act agc acc aag tcg gtc gtc cat tgg ttc cag att att cgg cac	1056
Phe Thr Ser Thr Lys Ser Val Val His Trp Phe Gln Ile Ile Arg His	
340 345 350	
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Arg Asn Phe Gln Phe Tyr Asp Asp Glu Ile His Ala Pro Leu Ser Ile	
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gtg gcc agt gag cga ttt tac aag ccg gtc aag tac ccg act aag aac	1152
Val Ala Ser Glu Arg Phe Tyr Lys Pro Val Lys Tyr Pro Thr Lys Asn	
370 375 380	
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Ile Lys Thr Pro Ile Val Leu Leu Tyr Gly Gly Ser Asp Ser Leu Val	
385 390 395 400	
gat atc aac gtg atg ttg tcc gag ctc cct cgc ggg acc gtg gcg aag	1248
Asp Ile Asn Val Met Leu Ser Glu Leu Pro Arg Gly Thr Val Ala Lys	
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Asp Gln Leu Val Phe Asn His Val Phe Glu Ala Leu Glu Arg Tyr Ser	
435 440 445	
tcg gag aat cag aaa ggg aca ttg atg gag aag gtt aat ggt gcc gcg	1392
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- <211> 470
- <212> PRT
- <213> Aspergillus niger

<400> 21
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21078WO.ST25.txt

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 65 70 75 80
 Asp Phe Thr Asp Ile Cys Ala Leu Phe Gly Tyr Glu Ala Glu Glu His
 85 90 95
 Ile Val Gln Thr Gly Asp Gly Tyr Leu Leu Gly Leu His Arg Leu Pro
 100 105 110
 Tyr Arg Lys Gly Glu Glu Gly Arg Lys Ile Asn Gln Gly Glu Gly Ser
 115 120 125
 Ile Lys Lys Lys Val Val Tyr Leu His His Gly Leu Met Met Cys Ser
 130 135 140
 Glu Val Trp Ile Cys Leu Ser Glu Glu Gln Arg Cys Leu Pro Phe Gln
 145 150 155 160
 Leu Val Glu Arg Gly Tyr Asp Val Trp Leu Gly Asn Asn Arg Gly Asn
 165 170 175
 Lys Tyr Ser Lys Lys Ser Val Lys His Ser Pro Leu Ser Asn Glu Phe
 180 185 190
 Trp Asp Phe Ser Ile Asp Gln Phe Ser Phe His Asp Ile Pro Asp Ser
 195 200 205
 Ile Lys Tyr Ile Leu Glu Val Thr Gly Gln Pro Ser Leu Ser Tyr Val
 210 215 220
 Gly Phe Ser Gln Gly Thr Ala Gln Ala Phe Ala Thr Leu Ser Ile His
 225 230 235 240
 Pro Leu Leu Asn Gln Lys Ile Asp Val Phe Val Ala Leu Ala Pro Ala
 245 250 255
 Met Ala Pro Thr Gly Leu Pro Asn His Leu Val Asp Ser Leu Met Lys
 260 265 270
 Ala Ser Pro Asn Phe Leu Phe Leu Leu Phe Gly Arg Arg Ser Ile Leu
 275 280 285
 Ser Ser Thr Thr Met Trp Gln Thr Ile Leu Tyr Pro Pro Ile Phe Val
 290 295 300
 Trp Ile Ile Asp Thr Ser Leu Arg Gly Leu Phe Asn Trp Arg Cys Lys
 305 310 315 320
 Asn Ile Ser Arg Trp Gln Lys Leu Ala Gly Tyr Leu His Leu Phe Ser
 325 330 335
 Phe Thr Ser Thr Lys Ser Val Val His Trp Phe Gln Ile Ile Arg His
 340 345 350
 Arg Asn Phe Gln Phe Tyr Asp Asp Glu Ile His Ala Pro Leu Ser Ile
 355 360 365
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	405		410	415
Glu Ile Pro Gln Tyr Glu	His Leu Asp Phe Leu	Trp Ala Arg Asp Val		
	420	425	430	
Asp Gln Leu Val Phe Asn	His Val Phe Glu Ala	Leu Glu Arg Tyr Ser		
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Ser Glu Asn Gln Lys Gly	Thr Leu Met Glu Lys	Val Asn Gly Ala Ala		
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- <210> 22
- <211> 3328
- <212> DNA
- <213> Aspergillus niger

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21078WO.ST25.txt

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- <210> 23
- <211> 1779
- <212> DNA
- <213> *Aspergillus niger*

- <220>
- <221> CDS
- <222> (1)..(1779)

21078WO.ST25.txt

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Leu Leu Phe Ala Gln Gln Leu Ala Ser His Pro Thr Glu Gln Ile Gln
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gcc att ctg gct ccg tgg gtc ccg gcc gca cta caa gat gtc gtg ctc      144
Ala Ile Leu Ala Pro Trp Val Pro Ala Ala Leu Gln Asp Val Val Leu
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Tyr Asn Arg Pro Arg Val Ile Ile Pro Gln Gly Thr Val Val Gly Thr
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Thr Leu Thr Asp Thr Leu Lys Ser Pro Val Asp Ala Phe Arg Gly Ile
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cca tac gca ttg cct cca att ggg gat aga cgg ttt cgc cgt gcg gag      288
Pro Tyr Ala Leu Pro Pro Ile Gly Asp Arg Arg Phe Arg Arg Ala Glu
85           90           95
gct gtc cat gcg acg gac gag att atc gat gct agt gaa ttc ggc cca      336
Ala Val His Ala Thr Asp Glu Ile Ile Asp Ala Ser Glu Phe Gly Pro
100          105          110
agg tgc cct gga aag cag ctc ttg aat cca aat gac ata ggt ggt gat      384
Arg Cys Pro Gly Lys Gln Leu Leu Asn Pro Asn Asp Ile Gly Gly Asp
115          120          125
gaa gac tgt ctc aca gtc aat gtc ttc cgg cct cat ggc gct cag gga      432
Glu Asp Cys Leu Thr Val Asn Val Phe Arg Pro His Gly Ala Gln Gly
130          135          140
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Lys Leu Pro Val Ala Val Tyr Val His Gly Gly Ala Tyr Asn Arg Gly
145          150          155          160
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Thr Ala Lys Tyr Pro Ala Ser Gly His Asn Thr Ala Ser Met Val Gly
165          170          175
tgg tcg gac gag ccc ttc gtt gca gtc agc ttc aac tac cgc atc ggc      576
Trp Ser Asp Glu Pro Phe Val Ala Val Ser Phe Asn Tyr Arg Ile Gly
180          185          190
gcc ctc ggc ttc ctc cca tcc acc cta acc gcc aaa gaa gga atc ctc      624
Ala Leu Gly Phe Leu Pro Ser Thr Leu Thr Ala Lys Glu Gly Ile Leu
195          200          205
aac cta ggc ctc cat gac cag atc ctc ctg ctg caa tgg gtc caa gaa      672
Asn Leu Gly Leu His Asp Gln Ile Leu Leu Leu Gln Trp Val Gln Glu
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Pro Pro Asn Thr Pro Leu Phe His Arg Ala Ile Ile Glu Ser Gly Ala
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gcc acc tcc cgc gcc gtc cac ccc tac aac gcc tcc ctc cac gaa tcc      864
Ala Thr Ser Arg Ala Val His Pro Tyr Asn Ala Ser Leu His Glu Ser
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caa ttc aca gac ttc ctc act gaa acg ggc tgc act aac ctc ccc gac      912
Gln Phe Thr Asp Phe Leu Thr Glu Thr Gly Cys Thr Asn Leu Pro Asp
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Thr Ala Ile Leu Pro Cys Leu Arg Ala Leu Pro Ser Ser Ala Ile Thr
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acc gcc tcc atc tcc gtc ttc gac aaa tac aac ccc tcc atc cgc tgg      1008
Thr Ala Ser Ile Ser Val Phe Asp Lys Tyr Asn Pro Ser Ile Arg Trp
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gcc ttc caa ccc gtc atc gac cac gag atc atc cac cgc cgg ccc atc      1056
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Phe Asn Ser Asn Glu Gly Thr Tyr Tyr Val Pro Arg Asn Leu Ser Leu
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Ser Glu Asp Phe Thr Ser Phe Phe Arg Thr Leu Leu Pro Ala Tyr Pro
385                390                395                400
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Glu Ser Asp Ile Gln Thr Ile Asp Glu Ile Tyr Pro Asp Pro Asn Val
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tat gct acg gcg tcg cca tac ctc gag aca agg ccg atc ccg agt cta      1296
Tyr Ala Thr Ala Ser Pro Tyr Leu Glu Thr Arg Pro Ile Pro Ser Leu
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Gly Arg Gln Phe Lys Arg Leu Glu Ala Ala Tyr Gly His Tyr Ala Tyr
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gcg tgt cca gta cgg cag acg gcg ggg ttt gtt gct aat gat gat ggt      1392
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Pro Ala Val Arg Asp Ile Ser Glu Ala Gln Arg Glu Val Ala Gly Leu	
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Phe His Ala Tyr Val Thr Ser Phe Val Val His Gly Asp Pro Asn Val	
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- <211> 592
- <212> PRT
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Ala Ile Leu Ala Pro Trp Val Pro Ala Ala Leu Gln Asp Val Val Leu
35 40 45
Tyr Asn Arg Pro Arg Val Ile Ile Pro Gln Gly Thr Val Val Gly Thr
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Thr Leu Thr Asp Thr Leu Lys Ser Pro Val Asp Ala Phe Arg Gly Ile
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435 440 445
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485 490 495
Pro Ala Val Arg Asp Ile Ser Glu Ala Gln Arg Glu Val Ala Gly Leu
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Phe His Ala Tyr Val Thr Ser Phe Val Val His Gly Asp Pro Asn Val
515 520 525
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Val Phe Trp Met His Gly Gly Glu Phe Ala Glu Gly Gly Thr Arg Asp	
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225 230 235 240
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 420 425 430

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145 150 155 160

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165 170 175

Phe Leu Tyr Ser Gln Glu Val Ala Asp Glu Gly Ser Ala Asn Leu Gly
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195 200 205

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275 280 285

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290 295 300

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Arg Ala Ser Gly Ile Val Asn Leu Gln Thr Gly Gln Phe Ala Lys Thr
325 330 335

Pro Leu Leu Ile Gly Thr Asn Phe Asp Glu Gly Thr Lys Tyr Ala Pro
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His Gly Tyr Asn Thr Thr Asp Gln Phe Val Ser Leu Val Gln Ala Asn
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Gly Thr Asn Tyr Thr Ser Ala Leu Thr Ile Ala Ser Leu Tyr Pro Asp
370 375 380

Asp Pro Ala Val Gly Ile Pro Gly Thr Leu Gln Gly Arg Pro Pro Pro
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Ser Tyr Gly Tyr Gln Trp Lys Arg Val Ala Ala Phe Leu Gly Asp Leu
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Leu Met His Ala Pro Arg Arg Val Thr Thr Gln Trp Leu Ala His Trp
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Asn Val Pro Ala Tyr Val Tyr His Trp Asn Val Met Thr Leu Gly Pro
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 Val Ser Ser Gln Asn Pro Thr Val Asp Leu Gly Tyr Thr Arg Tyr Lys
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Leu Phe Leu Asp Val Tyr Ala Pro Ser Ser Val Glu Ala Thr Thr Arg			
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Gln	Gln	Ala	Pro	Leu	Tyr	Leu	Tyr	Gly	Pro	Val	Val	Asp	Gly	Ser	Leu	
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Gln	Phe	Pro	Asn	Ile	Asn	Phe	Thr	His	Leu	Thr	Lys	Leu	Asn	Asp	Trp	
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Pro	Asn	Pro	Leu	Arg	Tyr	Pro	Gly	Ser	Pro	Glu	Trp	Lys	Thr	Trp	Ser	
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 65 70 75 80
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Tyr Leu Lys Glu Asn Gln Thr Arg Glu Phe Pro Asn Ser Ser Pro Tyr
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cgcgaggca tgatggagtc cggcgcggtg ttacctggca gtgccttga cctcacctgg 3240
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- <210> 32
- <211> 1695
- <212> DNA
- <213> Aspergillus niger

- <220>
- <221> CDS

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21078WO.ST25.txt

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agc ctg gca acc tgc acc aac cca gta gcc cag aca aag aac gga agt      96
Ser Leu Ala Thr Cys Thr Asn Pro Val Ala Gln Thr Lys Asn Gly Ser
20          25          30
tat tat ggt gtc tac atg cct cag tat aat gag gat tat ttt ctt gga      144
Tyr Tyr Gly Val Tyr Met Pro Gln Tyr Asn Glu Asp Tyr Phe Leu Gly
35          40          45
att cca ttt gct aag ccc ccg ttg gca cac ttg cgt tgg gcc aac ccc      192
Ile Pro Phe Ala Lys Pro Pro Leu Ala His Leu Arg Trp Ala Asn Pro
50          55          60
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Glu Ser Leu Asn Glu Ser Trp Ser Gly Leu Arg Pro Ala Thr Gly Tyr
65          70          75          80
gcg atg gaa tgt ata ggt tac ggc agt gat caa aaa ggt tat ctg cag      288
Ala Met Glu Cys Ile Gly Tyr Gly Ser Asp Gln Lys Gly Tyr Leu Gln
85          90          95
agc gag gac tgt ctc tac cta aac gtg gtc cgt ccc gct gaa tac gac      336
Ser Glu Asp Cys Leu Tyr Leu Asn Val Val Arg Pro Ala Glu Tyr Asp
100         105         110
aat gcc agt ctt cca gtc ctt gta tgg att cat ggc ggt ggc ttc gca      384
Asn Ala Ser Leu Pro Val Leu Val Trp Ile His Gly Gly Gly Phe Ala
115         120         125
caa ggc ggc act ccc gac ctt cga tac aat ctt aca ttt att gtt gaa      432
Gln Gly Gly Thr Pro Asp Leu Arg Tyr Asn Leu Thr Phe Ile Val Glu
130         135         140
cac tcg gtc aat atc ggc cag cca att atc gca gtg agc gtt gcc tat      480
His Ser Val Asn Ile Gly Gln Pro Ile Ile Ala Val Ser Val Ala Tyr
145         150         155         160
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cgt ctc ggt cct tgg ggt ttc ttc aat ggg gtc gag ctc gcc aat gag      528
Arg Leu Gly Pro Trp Gly Phe Phe Asn Gly Val Glu Leu Ala Asn Glu
165         170         175
gga tcg tta aat ctc ggg ctg aag gac cag cgc ttg gcc ctg cat tgg      576
Gly Ser Leu Asn Leu Gly Leu Lys Asp Gln Arg Leu Ala Leu His Trp
180         185         190
gtg aaa gag aac att gca ggt ttc ggt ggc gac cct agt aaa gtc gtg      624
Val Lys Glu Asn Ile Ala Gly Phe Gly Gly Asp Pro Ser Lys Val Val
195         200         205
att tac gga caa agt gcc ggc tcc gaa agc gtg gga tac caa atc cgc      672
Ile Tyr Gly Gln Ser Ala Gly Ser Glu Ser Val Gly Tyr Gln Ile Arg

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210	215	220	
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225	230	235	240
tcc ggc gcg gtg tta cct ggc agt gcc ttg aac ctc acc tgg aca tat			768
Ser Gly Ala Val Leu Pro Gly Ser Ala Leu Asn Leu Thr Trp Thr Tyr			
	245	250	255
gag cct tgg ttc cag caa ata gca gac gag gca gga tgt tcc cag acc			816
Glu Pro Trp Phe Gln Gln Ile Ala Asp Glu Ala Gly Cys Ser Gln Thr			
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acc cgc aaa ctg gac tgt cta cgc cgc acg ccc ttc aca gtc cta aac			864
Thr Arg Lys Leu Asp Cys Leu Arg Arg Thr Pro Phe Thr Val Leu Asn			
	275	280	285
aac att ctg aac acc acc gcc aac gac acg acg cct tac aac tgg agg			912
Asn Ile Leu Asn Thr Thr Ala Asn Asp Thr Thr Pro Tyr Asn Trp Arg			
	290	295	300
ccc aca gtg gac ggt gac ttc gta gcg cga tat ccc agc gag caa ctc			960
Pro Thr Val Asp Gly Asp Phe Val Ala Arg Tyr Pro Ser Glu Gln Leu			
305	310	315	320
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Asp Thr Gly Asp Phe Val Lys Val Pro Ile Ile Ile Gly Tyr Thr Thr			
	325	330	335
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Asp Glu Gly Thr Thr Glu Cys Pro Glu Pro Val Asn Thr Thr Ala Glu			
	340	345	350
tta aaa gaa tac ctc agc tca aca aca acc tac ggc tgg gcc ctc gac			1104
Leu Lys Glu Tyr Leu Ser Ser Thr Thr Thr Tyr Gly Trp Ala Leu Asp			
	355	360	365
tca cag gta gta tcc tcg ctc ctg gac ctc tac ccc aac acc acc tcc			1152
Ser Gln Val Val Ser Ser Leu Leu Asp Leu Tyr Pro Asn Thr Thr Ser			
	370	375	380
ttc ggc atc cca tca tcc gaa gaa ctc ggc ggc aac gtc acc ttc cca			1200
Phe Gly Ile Pro Ser Ser Glu Glu Leu Gly Gly Asn Val Thr Phe Pro			
385	390	395	400
cag ccc tac ggc gcc gca ttc cgc cag acg gca gca tac tac ggc gac			1248
Gln Pro Tyr Gly Ala Ala Phe Arg Gln Thr Ala Ala Tyr Tyr Gly Asp			
	405	410	415
gcc cag ttc ata gcc gcg acg cgc tac acc tgt gag cta tgg gcg gca			1296
Ala Gln Phe Ile Ala Ala Thr Arg Tyr Thr Cys Glu Leu Trp Ala Ala			
	420	425	430
cat aac ctg aca gca tat tgc tac cga ttc aac acc aag aca gac gat			1344
His Asn Leu Thr Ala Tyr Cys Tyr Arg Phe Asn Thr Lys Thr Asp Asp			
	435	440	445
tac aac agg gaa gaa ggc gtg gcg cat ttc tcg gac gtg atc ttc atc			1392

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 Phe Asn Asn Leu Asn Gly Tyr Gly Phe Ser Pro Asn Pro Phe Thr Asn
 465 470 475 480
 gct cca gag agc tat act gag ctt agc tac ctc atg tcc ggc tcg tgg 1488
 Ala Pro Glu Ser Tyr Thr Glu Leu Ser Tyr Leu Met Ser Gly Ser Trp
 485 490 495
 atc agc ttc act aat agt ctg gat cct aat aag tgg act ggt cgc gga 1536
 Ile Ser Phe Thr Asn Ser Leu Asp Pro Asn Lys Trp Thr Gly Arg Gly
 500 505 510
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 Arg Asn Ala Thr Lys Thr Glu Asn Trp Pro Val Tyr Asp Leu Glu Asn
 515 520 525
 ccc ttg agt atg atc tgg gat gcg aat gtc act tcg tat gcg gcg ccg 1632
 Pro Leu Ser Met Ile Trp Asp Ala Asn Val Thr Ser Tyr Ala Ala Pro
 530 535 540
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 Ala Tyr Gln Arg

- <210> 33
- <211> 564
- <212> PRT
- <213> Aspergillus niger

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 35 40 45
 Ile Pro Phe Ala Lys Pro Pro Leu Ala His Leu Arg Trp Ala Asn Pro
 50 55 60
 Glu Ser Leu Asn Glu Ser Trp Ser Gly Leu Arg Pro Ala Thr Gly Tyr
 65 70 75 80
 Ala Met Glu Cys Ile Gly Tyr Gly Ser Asp Gln Lys Gly Tyr Leu Gln
 85 90 95
 Ser Glu Asp Cys Leu Tyr Leu Asn Val Val Arg Pro Ala Glu Tyr Asp
 100 105 110

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21078WO.ST25.txt

Asn Ala Ser Leu Pro Val Leu Val Trp Ile His Gly Gly Gly Phe Ala
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130 135 140

His Ser Val Asn Ile Gly Gln Pro Ile Ile Ala Val Ser Val Ala Tyr
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Arg Leu Gly Pro Trp Gly Phe Phe Asn Gly Val Glu Leu Ala Asn Glu
165 170 175

Gly Ser Leu Asn Leu Gly Leu Lys Asp Gln Arg Leu Ala Leu His Trp
180 185 190

Val Lys Glu Asn Ile Ala Gly Phe Gly Gly Asp Pro Ser Lys Val Val
195 200 205

Ile Tyr Gly Gln Ser Ala Gly Ser Glu Ser Val Gly Tyr Gln Ile Arg
210 215 220

Ala Tyr Asn Gly Arg Asp Asp Gly Leu Phe Arg Gly Gly Met Met Glu
225 230 235 240

Ser Gly Ala Val Leu Pro Gly Ser Ala Leu Asn Leu Thr Trp Thr Tyr
245 250 255

Glu Pro Trp Phe Gln Gln Ile Ala Asp Glu Ala Gly Cys Ser Gln Thr
260 265 270

Thr Arg Lys Leu Asp Cys Leu Arg Arg Thr Pro Phe Thr Val Leu Asn
275 280 285

Asn Ile Leu Asn Thr Thr Ala Asn Asp Thr Thr Pro Tyr Asn Trp Arg
290 295 300

Pro Thr Val Asp Gly Asp Phe Val Ala Arg Tyr Pro Ser Glu Gln Leu
305 310 315 320

Asp Thr Gly Asp Phe Val Lys Val Pro Ile Ile Ile Gly Tyr Thr Thr
325 330 335

Asp Glu Gly Thr Thr Glu Cys Pro Glu Pro Val Asn Thr Thr Ala Glu
340 345 350

Leu Lys Glu Tyr Leu Ser Ser Thr Thr Thr Tyr Gly Trp Ala Leu Asp
355 360 365

~~Ser Gln Val Val Ser Ser Leu Leu Asp Leu Tyr Pro Asn Thr Thr Ser~~
370 375 380

Phe Gly Ile Pro Ser Ser Glu Glu Leu Gly Gly Asn Val Thr Phe Pro
385 390 395 400

Gln Pro Tyr Gly Ala Ala Phe Arg Gln Thr Ala Ala Tyr Tyr Gly Asp
405 410 415

Ala Gln Phe Ile Ala Ala Thr Arg Tyr Thr Cys Glu Leu Trp Ala Ala
420 425 430

His Asn Leu Thr Ala Tyr Cys Tyr Arg Phe Asn Thr Lys Thr Asp Asp
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Tyr Asn Arg Glu Glu Gly Val Ala His Phe Ser Asp Val Ile Phe Ile
450 455 460

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21078WO.ST25.txt

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 465 470 475 480
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 485 490 495
 Ile Ser Phe Thr Asn Ser Leu Asp Pro Asn Lys Trp Thr Gly Arg Gly
 500 505 510
 Arg Asn Ala Thr Lys Thr Glu Asn Trp Pro Val Tyr Asp Leu Glu Asn
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 Pro Leu Ser Met Ile Trp Asp Ala Asn Val Thr Ser Tyr Ala Ala Pro
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- <210> 34
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- <212> DNA
- <213> Aspergillus niger

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 <213> Aspergillus niger

<220>
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 Gln Phe Gly Thr Gly Ser Thr Ala Asn Glu Leu Glu Gln Gly Ser Cys
 35 40 45
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 50 55 60
 atg gcc acc gta atc gcc ccc cct ctc tgc gac aac ctg aaa tcc aaa 240
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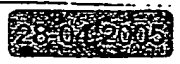
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      85                      90                      95
gcc gga ctc gtg cag aat gcc ctg ccc cag aac acc gat ccg ggg agt      336
Ala Gly Leu Val Gln Asn Ala Leu Pro Gln Asn Thr Asp Pro Gly Ser
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atc tcc gcc gcg aag cag atg ttc gag gag gcg aat tcg aag tgt ccc      384
Ile Ser Ala Ala Lys Gln Met Phe Glu Glu Ala Asn Ser Lys Cys Pro
      115                     120                     125
aat act aag att gtt gcg ggt ggt tat agt caa gga agc gct gtg att      432
Asn Thr Lys Ile Val Ala Gly Gly Tyr Ser Gln Gly Ser Ala Val Ile
      130                     135                     140
gac aac gcc gtg caa gaa ctc agc acc acc gtg aaa gac caa gtg aag      480
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      145                     150                     155                     160
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Gly Val Val Leu Phe Gly Phe Thr Arg Asn Val Gln Asp His Gly Gln
      165                     170                     175
atc cct aat tac cct aag gat gac gtg aag gtt tat tgt gcc gtg ggc      576
Ile Pro Asn Tyr Pro Lys Asp Asp Val Lys Val Tyr Cys Ala Val Gly
      180                     185                     190
gat ctg gtc tgt gat gat acg ttg gtt gtt acg gcg atg cat ctg acg      624
Asp Leu Val Cys Asp Asp Thr Leu Val Val Thr Ala Met His Leu Thr
      195                     200                     205
tat ggc atg gat gcg ggt gat gcg gcg agc ttt ttg gcc gag aag gtg      672
Tyr Gly Met Asp Ala Gly Asp Ala Ala Ser Phe Leu Ala Glu Lys Val
      210                     215                     220
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      225                     230                     235                     240
agt tca tct gct gcg ggg acg tcg tcg tcg ggg ttg tcg gga ctg tct      768
Ser Ser Ser Ala Ala Gly Thr Ser Ser Ser Gly Leu Ser Gly Leu Ser
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- <210> 36
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- <212> PRT
- <213> Aspergillus niger

- <400> 36



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65 70 75 80
Leu Gly Ser Asp Lys Val Ala Cys Gln Gly Val Gly Gly Gln Tyr Ser
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Gly Val Val Leu Phe Gly Phe Thr Arg Asn Val Gln Asp His Gly Gln
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Ile Pro Asn Tyr Pro Lys Asp Asp Val Lys Val Tyr Cys Ala Val Gly
180 185 190
Asp Leu Val Cys Asp Asp Thr Leu Val Val Thr Ala Met His Leu Thr
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Tyr Gly Met Asp Ala Gly Asp Ala Ala Ser Phe Leu Ala Glu Lys Val
210 215 220
Gln Ser Ser Ser Ser Ser Thr Thr Ser Ser Ser Ser Asp Ala Ala Ser
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- <210> 37
- <211> 2981
- <212> DNA
- <213> Aspergillus niger

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 515 520 525
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 530 535 540
 Arg Lys Glu Gly Ile Asp Tyr Ile Asn Ser Val Ala Asn Ala Tyr Trp
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